Identification of blast resistance genes in IRRI-bred rice varieties by segregation analysis based on a differential system

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Abstract

A differential system for rice blast disease consists of standard blast isolates (Pyricularia oryzae Cavara..) and differential rice varieties (Oryza sativa L.) with known resistance genes. Monogenic lines for blast resistance have been newly developed at the International Rice Research Institute (IRRI) as international differential varieties and differential blast isolates from the Philippines were selected. Using a differential system based on the gene-for-gene theory, blast resistance genes were estimated in the IRRI-bred elite rice varieties. At least seven kinds of resistance genes --Pi20(t), Pita, Pik* (one of the Pik allele genes except Pik-s), Pik-s, Pib, Piz-t, and Pii or Pi3 -- were estimated in 42 rice varieties on the basis of reaction patterns to 14 standard blast isolates. The reaction patterns of the IRRI varieties were compared with those of the blast monogenic lines. To confirm this gene estimation, genetic analysis was done using segregating populations derived from crosses between the IRRI varieties and a susceptible Indica-type variety, CO 39. The BC_1F_2 populations (with CO 39 as recurrent parent) were segregated for reaction to the specific standard isolates. Furthermore, the estimated genes were confirmed by allelism test against the blast resistance monogenic lines. As a result of the segregation analysis of 10 of 42 IRRI-bred varieties, seven genes -- Pi20(t), Pita, Pik*, Pia, Pib, Piks -- and Piz-t, were identified. Genes Pia, Pib, Pik-s, and Piz-t were not estimated by reaction patterns to the blast isolates but identified by genetic analysis in some varieties. The effectiveness of the differential system, which is based on conventional methods and which does not require advanced facilities, is discussed as a fundamental tool to provide essential information to develop breeding programs for blast resistance.

Keywords: rice (Oryza sativa L.), blast (Pyricularia oryzae Cavara), differential system, genetic analysis, resistance, IRRI variety

Introduction

Blast disease, caused by *Pyricularia oryzae* Cavara is a serious disease of rice (*Oryza sativa* L.) worldwide. The use of host resistance is one of the most economical and effective means of controlling blast disease. Resistance to blast disease is governed by a gene-for-gene relationship between resistance gene in the host and avirulence gene in the blast pathogen (Kiyosawa, 1972; Silue et. al., 1992). Genetic studies performed in this context led to the identification of many blast resistance genes. Genetic studies in Indica-type varieties have been limited because the extremely changeable virulence of the blast fungus (Ou, 1985; Bonman et al., 1986) and the presence of several resistance genes in Indica-type varieties' genetic background (Mackill et al.; 1985; Yu et al., 1987) made these studies complicated.

The limitation in genetic studies on blast resistance could also be attributed to a lack of suitable differential system for blast resistance genes in the tropics. The 12 Japonica-type differential varieties (DVs) for blast resistance were selected by Yamada et al. (1976) and Kiyosawa (1981), but those DVs are known to carry additional gene(s) showing resistance to tropical blast isolates, which masked the targeted gene's reaction (Noda et al., 1999). To address this limitation, Yanoria et al. (2000) tried to clarify the pathogenicities of blast isolates from the Philippines using DVs and lines. Several blast isolates with distinct pathogenicities were selected and studied in detail by Tsunematsu et al. (2000) using a set of monogenic lines that contains 23 kinds of single resistance genes with the genetic background of a Chinese variety Lijiang-xin-tuan-heigu (LTH).

In South and Southeast Asia, rice varieties bred in IRRI are widely grown and account for more than 80% of total rice production in this region (Khush, 1990). And these IRRI-bred varieties have been used for breeding materials in many countries of Tropical region. Using the differential system consisting of newly developed monogenic lines and differential blast isolates from the Philippines, kinds of blast resistance genes in IRRI-bred varieties were identified. In this paper, methodologies of gene identification are explained following 3 steps with special emphasis on a case of IR64, which were distributed and cultivated the most widely and was still one of leading varieties in tropical region At first, kind of blast resistance gene was estimated by reaction patterns to differential blast isolates (I). To confirm the gene estimation, segregation analysis using backcross progenies with a susceptible variety (II) and allelism test with DVs were conducted (III).

I. Gene estimation based on reaction pattern to differential blast isolates

Materials and methods

1. Rice plant materials

A total of 42 IRRI-bred rice varieties were used for blast resistance gene estimation (Table 1). The widely grown varieties, IR8, IR24, IR36, IR64 and IR72 were included. Eight varieties, designated as PSBRc by the Philippine Seed Board, which are IRRI developed varieties, were also included.

Nine monogenic lines, IRBL20-IR24 (*Pi20*(t)), IRBLta-CP1 (*Pita*), IRBLk-K (*Pik*), IRBLks-F5 (*Piks*), IRBLa-A (*Pia*), IRBLb-B (*Pib*), IRBLzt-T (*Pizt*), IRBLi-F5 (*Pii*), and IRBL3-CP4 (*Pi3*) were tested for comparison.

2. Inoculation and evaluation of resistance

Pregerminated seeds of each variety or population were sown on soil in a 26×35 cm plastic tray. Ten grams of ammonium sulfate was applied to each tray as basal fertilizer and 1 g was added 1 wk before inoculation. In all cases, susceptible cultivars CO39 and LTH were included in each tray to check the success of inoculation and the virulence of blast isolates used.

Fourteen blast isolates with known avirulence (Yanoria et al., 2000, Telebanco-Yanoria et al. 2007) from the Philippines were used for inoculation. Seedlings at the four-leaf stage were sprayed with 40-50 ml spore suspension per tray adjusted to 10^5 spores ml⁻¹. Trays were placed inside the wet jute sacks for 18-24 h, and transferred to an air-conditioned glasshouse room at 23 ± 3 °C/ 30 ± 5 °C night and day temperature. Blast incidence on each seedling was examined 6-7 d after inoculation. The reaction was classified into 0-5 scales with slight modifications as described previously by Mackill and Bonman (1992). Seedlings rated 0-2 were considered resistant (R), 3 was moderate resistant (M), and those rated 4-5, susceptible (S).

3. Resistance gene estimation

Gene(s) were estimated by comparing reaction patterns to the differential blast isolates of IRRI varieties with those of blast monogenic lines (Table 2). Using the Philippine blast isolates, 4 alleles of *Pik* loci (*Pik*, *Pik-h*, *Pik-m*, and *Pik-p*) can not be differentiated because all the monogenic lines with these genes showed the same reaction pattern (Tsunematsu et al. 2000). Therefore, those alleles were designated tentatively as *Pik** in this paper.

Although total 14 differential isolates were involved in this study, reactions to 3 distinct isolates, M36-1-3-10-1, IK81-25, and PO6-6, were considered for grouping of varieties and primary gene estimation as a first step. As a second step, reaction patterns to the other 11 isolates were also taken into consideration for gene estimation (Table 2).

Results and discussion

1. Gene estimation in IR64

IR64 was resistant to 12 (M36-1-3-10-1, IK81-25, IK81-3, BN111, V850256, M39-1-3-8-1, M64-1-3-9-1, M39-1-2-21-2, BN209, V850196, B90002, and C923-49) out of 14 standard blast isolates tested (Table 3). Among the 9 monogenic lines tested, 6 lines (IRBL20-IR24, IRBLta-K1, IRBLb-B, IRBLa-A, IRBLks-F5, and IRBLzt-T) were resistant to the blast isolates which were avirulent to IR64. These results suggested that 6 blast resistant genes (Pi20(t), Pita, Pib, Pia, Pik-s, and Piz-t) which were harbored in the respective monogenic lines were estimated to exist in the genetic background of IR64. Three genes, Pi20(t), Pita, and Pib showed relatively wide spectrum of resistance and covered the resistance of the other 3 genes, Pia, Pik-s, and Piz-t. Therefore, the latter 3 were not estimated directly but possible to be harbored in the genetic background.

The other 3 monogenic lines, IRBLk-Ka, IRBLi-F5, and IRBL3-CP4 were resistant to a blast isolate PO6-6 which was virulent to IR64 (Table 3). This suggested that 3 resistant genes, *Pik*, *Pii*, and *Pi3* were not harbored in the genetic background of IR64.

2. Gene estimation in IRRI-bred varieties

IRRI-bred varieties showed various patterns to the differential blast isolates. The reaction patterns of these varieties were compared with those of DVs (Table 4). These varieties were classified into 8 variety groups (VG) based on reaction patterns to three distinct blast isolates, M36-1-3-10-1, IK81-25, and PO6-6 (Table 4). These 3 isolates showed incompatibility with 3 resistance genes, *Pi20*(t) resistant to M36-1-3-10-1, *Pita* resistant to IK81-25, and *Pik* resistant to PO6-6. From these specific reactions, presence or absence of these 3 genes was estimated in the varieties for each VG.

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Variety	Year of release	Ecosystem suited	Line designation	Cross combination
IR 5	1967	Irrigated	IR5-47-2	Peta/ tangkai Rotan
IR 8	1966	Irrigated	IR8-288-2	Pata/Dee-geo-woo-gen
IR20	1969	Irrigated	IR523-E576	IR262-24-3/TKM6
IR22	1969	Irrigated	IR579-160-2	IR8/Tadukan
IR24	1971	Irrigated	IR661-1-140-3	IR8/IR127-2-2
IR26	1973	Irrigated	IR1541-102-7	IR24/TKM6
IR 28	1974	Irrigated	IR2061-214-3-8-2	IR833///IR1561-149-1//IR24*4/O. nivara
IR 29	1974	Irrigated	IR2061-464-4-14-1	IR833///IR1561-149-1//IR24*4/O. nivara
IR 30	1974	Irrigated	IR2153-159-1-4	IR1541-102-6-3//IR24*4/O. nivara
IR32	1975	Irrigated	IR2070-747-6-3-2	IR20*2/O.nivara//CR94-13
IR34	1975	Irrigated	IR2061-213-2-17	IR833-6-2-1-1//IR1561-149-1//IR24*4/O.nivara
IR36	1976	Irrigated	IR2071-625-1-252	IR1561-228-1-2/IR737//CR94-13
IR38	1976	Irrigated	IR2070-423-2-5-6	IR20*2/O.nivara//CR94-13
IR 40	1977	Irrigated	IR2070-414-3-9	IR20*2/O.nivara//CR94-13
IR42	1977	Irrigated	IR2071-586-5-6-3	IR1561-228-1-2/IR737//CR94-13
IR 43	1978	Irrigated	IR1529-430-3	IR305-3-17-1-3/IR661-1-140-3
IR 44	1978	Irrigated	IR2863-38-1-2	IR1529-680-3/CR94-13//IR480-5-9-3
IR 45	1978	Irrigated	IR2035-242-1	IR1416-128-5/IR1364-37-3-1//IR1824-1
IR 46	1978	Irrigated	IR2058-78-1-3-2	IR1416-128-5/IR1364-37-3-1//IR1366-120-3-1/IR1539-111
IR 48	1979	Irrigated	IR4570-83-3-3	IR1702-74-3-2/IR1721-11-6-8-3//IR2055-481-2
IR 50	1979	Irrigated	IR9224-117-2-3-3-2	IR2153-14-1-6-2/IR28//IR36
IR 52	1980	Irrigated	IR5853-118-5	Nam Sa-gui 19/IR2071-88//IR2061-214-3-6-20
IR 54	1980	Irrigated	IR5853-162-1-2-3	Nam Sa-gui 19/IR2071-88//IR2061-214-3-6-20
IR 56	1982	Irrigated	IR13429-109-2-2-1	IR4432-53-33/PTB33//IR36
IR 58	1983	Irrigated	IR9752-71-3-2	IR28/Kwang-chang-ai//IR36
IR 60	1983	Irrigated	IR13429-299-2-1-3	IR4432-53-33/PTB33//IR36
IR 62	1984	Irrigated	IR13525-43-2-3-1-3-2	PTB33/IR30//IR36
IR 64	1985	Irrigated	IR18348-36-3-3	IR5657-33-2-1/IR2061-465-1-5-5
IR 65	1985	Irrigated	IR21015-196-3-1-3	Batatais/IR36//IR52
IR 66	1987	Irrigated	IR32307-107-3-2-2	IR13240-108-2-2-3/IR9129-209-2-2-2-1
IR 68	1988	Irrigated	IR28224-3-2-3-2	IR19660-73-4/IR2415-90-4-3-2//IR54
IR 70	1988	Irrigated	IR28228-12-3-1-1-2	IR19660-73-4/IR54//IR9828-36-3
IR 72	1988	Irrigated	IR35366-90-3-2-1-2	IR19661-9-2-3/IR1576-199-3-3//IR9129-209-2-2-2-1
IR74	1988	Irrigated	IR32453-20-3-2-2	IR19661-131-1-2/IR15795-199-3-3
PSBRc 1	1990	Upland	IR10147-113-5-1-1-5	KN-1B-361-1-8-6/IR1750-F5B-3//BPI76*9/Dawn
PSBRc 2	1991	Irrigated	IR32809-26-3-3	IR4215-301-2-2-6/BG90-2//IR19661-131-1-2
PSBRc 4	1991	Irrigated	IR41985-111-3-2-2	IR4547-4-1-2/IR1905-81-3-1//IR25621-94-3-2
PSBRc 10	1992	Irrigated	IR50404-57-2-2-3	IR33021-39-2-2/IR32429-47-3-2-2
PSBRc 18	1992	Irrigated	IR51672-62-2-1-1-2-3	IR24594-204-1-3-2-6-2/IR28222-9-2-2-2-2
PSBRc 20	1994	Irrigated	IR57301-195-3-3	IR35293-125-3-2-3/IR32429-47-3-2-2/PSBRc4
PSBRc 28	1995	Irrigated	IR56381-139-2-2	IR28239-94-2-3-6-2/IR64
PSBRc 30	1995	Irrigated	IR58099-41-2-3	IR72/IR24632-34-2
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Table 1. List of IRRI-bred rice varieties used for identifing the blast resistance genes

IRRI (1995) and Peng and Khush (2003) were modified.

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				Stand	lsrd di	fferentia	l blast	isolates	from the	e Philipp	pines			
Monogenic lines as differential varieties (resistance gene)	M36-1-3-10-1	IK81-25	PO6-6	IK81-3	BN111	Ca89	V850256	M39-1-3-8-1	M64-1-3-9-1	M39-1-2-21-2	BN209	V850196	B90002	C923-49
IRBL20-IR24 (<i>Pi20</i> (t))	R	S	S	S	R	S	S	S	S	R	S	S	R	R
IRBLta-CP1 (Pita)	S	R	S	R	S	S	R	R	R-M	S	S	R	R	S
IRBLk-Ka (Pik)	S	S	R	R	R	R	R	S	S	S	R	R	R	R
IRBLks-F5 (Pik-s)	S	S	S	S	S	S	S	S	S	S	S	R	S	S
IRBLa-A (Pia)	S	S	S	S	S	S	S	S	S	S	S	S	R	R
IRBLb-B (Pib)	S	S	S	S	S	S	S	S	S	S	R-M	S	R	R
IRBLzt-T (Piz-t)	S	S	S	S	S	S	S	R	R	S	S	S	R	R
IRBLi-F5 (Pii)	S	R-M	R	R-M	R	S	S	М	S	R-M	S	R	S	S
IRBL3-CP4 (Pi3(t))	S	R	R	М	R	S	S	R-M	S	R	S	R	S	S

Table 2. Reaction pattern of monoginic lines to standard blast isolates from the Philippines

Monogenic lines were developed as a set of diffrential varieties by Tsunematsu et al. 2000

Standard diffential blast isolates from the Philippines were selected by Telebanco-Yanoria et al. 2008.

					Standar	d diffe	erential	blast	isolates	from t	he Phil	lippines			
(Resistance	e genotype gene harbored in the c background)	M36-1-3-10-1	IK81-25	PO6-6	IK81-3	BN111	Ca89	V850256	M39-1-3-8-1	M64-1-3-9-1	M39-1-2-21-2	BN209	V850196	B90002	C923-49
IR64	(<i>Pi20</i> (t), <i>Pita</i> , <i>Pib</i>)*	R	R	S	R	R	S	R	R	R	R	R	R	R	R
	(Pia, Pik-s, Piz-t)**	ĸ	R	0	R	R	5	R	IX.	R	К	R	ĸ	R	R
IRBL20-IR2	4 (<i>Pi20</i> (t))	R	S	S	S	R	S	S	S	S	R	S	S	R	R
IRBLta-K1	(Pita)	S	R	S	R	S	S	R	R	R-M	S	S	R	R	S
IRBLb-B	(Pib)	S	S	S	S	S	S	S	S	S	S	R-M	S	R	R
IRBLa-A	(Pia)	S	S	S	S	S	S	S	S	S	S	S	S	R	R
IRBLks-F5	(Pik-s)	S	S	S	S	S	S	S	S	S	S	S	R	S	S
IRBLzt-T	(Piz-t)	S	S	S	S	S	S	S	R	R	S	S	S	R	R
IRBLk-Ka	(Pik)	S	S	R	R	R	R	R	S	S	S	R	R	R	R
IRBLi-F5	(Pii)	S	R-M	R	R-M	R	S	S	М	S	R-M	S	R	S	S
IRBL3-CP4	(<i>Pi3</i> (t))	S	R	R	М	R	S	S	R-M	S	R	S	R	S	S

Table 3. Blast resistance gene estimation in IR64 based on the differential system

S; susceptible, M; moderate resistant, and R; resistant.

*Estmated resistatce genes harboried in the IR64 genetic background, based on the reaction patterns to standard differential blast isolates from the Philippines

** Possible resistance gene harbored in the genetic background. These reaction of resistance genes are masked by the estimated genes which show the wide spectrums to the blast isolates

Three genes, *Pik*, *Pii*, and *Pi3*(t) are not harbored in the genetic background of IR64. Reaction patterns of the monogenic lines do not agree with that of IR64.

Variety Group and	Reaction to s	tandard blast isol	ate ¹⁾	Estimated gene(s) harboring in the genetic					
differential variety	M36-1-3-10-1	IK81-25	PO6-6	background					
IRBL20-IR24	R	S	S	<i>Pi20</i> (t)	-	-			
IRBLta-CP1	S	R	S	-	Pita	-			
IRBLk-K	S	S	R	-	-	Pik			
VG1	S	S	S	-	-	-			
VG2	R	S	S	<i>Pi20</i> (t)	-	-			
VG3	S	R	S	-	Pita	-			
VG4	М	S	R	-	-	Pik			
VG5	R	S	R	<i>Pi20</i> (t)	-	Pik-			
VG6	S	R	R	-	Pita	Pik			
VG7	R	R	S	<i>Pi20</i> (t)	Pita	-			
VG8	R	R	R	<i>Pi20(</i> t)	Pita	Pik			

Table 4. Variety classification based on reaction pattern to 3 selected blast isolates

S; susceptible, M; moderate resistant, and R; resistant.

1) Blast isolates were selcted from the Philippines by Telebanco-Yanoria et al. 2008

Five IRRI varieties, IR20, IR28, IR30, IR45, and IR66, were susceptible to all 3 selected isolates and classified as VG1a (Table 4). None of the Pi20(t), Pita, or Pik* were estimated in these varieties. These five varieties showed resistance to 4 isolates, BN209, V850196, B90002, and C923-49 (Table 5). These results can be explained by presence of Pib and Pik-s in VG1 (Table 2 and 5). Two varieties, IR29 and IR34 in VG1b showed resistance to 4 more isolates, IK81-25, IK81-3, M39-1-3-8-1, and M64-1-3-9-1. The resistant reactions to M39-1-3-8-1, and M64-1-3-9-1 were attributed to the presence of Piz-t in these 2 varieties, while those to the other 2 isolates, IK81-25 and IK81-3 were to unknown gene(s). From these reaction patterns, VG1 was further classified into 2 subgroups; i.e. VG1a (IR20, IR28, IR30, IR45, and IR66 estimated with Pib and Pik-s) and VG1b (IR29 and IR34 with *Pib*, *Pik-s*, *Piz-t* and unknown gene).

Seven varieties, IR8, IR22, IR24, IR26, IR43, PSBRc2, and PSBRc30, showed resistance to a blast isolate M36-1-3-10-1 and were classified as VG2 (Table 4). VG2 was estimated to harbor Pi20(t), because this gene is incompatible with the isolate M36-1-3-10. Resistance genes, *Pita* or *Pik** were thought to be absent in VG2, because these varieties was susceptible to both IK81-25 and PO6-6 which are avirulent to these genes. All 7 varieties in this group also showed resistance to 3 isolates, BN209, B90002, and C923-49, which were avirulent to *Pib* (Table 2 and 5). These results suggested that varieties in VG2 were likely to harbor *Pi20*(t) and *Pib*.

Furthermore, VG2 was divided into 3 subgroups,

VG2a, VG2b, and VG2c based on differences in reaction to 3 isolates, M39-1-3-8-1, M64-1-3-9-1, and V850196 (Table 5). The varieties in subgroups, VG2a and VG2c showed resistance to V850196 whereas VG2b showed moderate resistance. Resistance to this isolated was estimated to be controlled by a resistance gene *Pik-s*, because *Pita* nor *Pik** was not estimated in VG2. The moderate resistance to this isolate of VG2b containing only one variety IR43 could be attributed to the presence of an unknown gene. VG2b was resistant to M39-1-3-8-1 and moderately resistant to M64-1-3-9-1. This reaction pattern could not be explained by resistance gene in the DVs used in this study. VG2c which included only one variety, PSBRc2, was resistant to M39-1-3-8-1 and M64-1-3-9-1 likely because of the presence of *Piz-t*.

Among the IRRI varieties tested, many of them were resistant to IK81-25 and susceptible to both M36-1-3-10 and PO6-6 (Table 4). From this reaction pattern, 17 varieties, IR5, IR32, IR36, IR38, IR40, IR42, IR44, IR50, IR52, IR54, IR58, IR60, IR62, IR65, IR68, IR72, and PSBRc4, were classified as VG3 and considered to harbor Pita and not to harbor Pi20(t) or Pik*. Resistance to IK81-3, V850256, M39-1-3-8-1, M64-1-3-9-1, V850196, and B90002 were considered to be attribution of Pita (Table 2 and 5). VG3 varieties were also resistant to blast isolates BN209 and C923-49 which were avirulent to Pib. Therefore, VG3 were estimated to harbor Pita and Pib. The presence of Pita and Pib did not contradicted to susceptible reaction to PO6-6, M36-1-3-10-1, BN111, Ca89, and M39-1-2-21-2.

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	netic		ı		1	ı	ı	Pix		ı	Pix	,	1	,	<i>Pii</i> or <i>Pi3</i> (t)	
	Resistance genes estimated and harboring in the genetic background	ı	ı	·		Piz-t	1		Piz-t	ı	ı	1	ı		-	
	urboring	ī	ı	ı	Pib	Pib	Pib	Pib	Pib	Pib	ī	ı	ı	Pib	Pib	
	nated and ha background	ı	ı	ı	Pik-s	Pik-s										
	estimate bac	ı	I	Pik	1	I	ı	I	ı		Pik^*	Pik*	Pik*	ı	ı	
	ce genes	ī	Pita	ı	1	ı	ı	ı	ı	Pita	ī	ı	Pita	Pita	Pita	l Pik-p.
	Resistan	<i>Pi20</i> (t)	ı	ı			<i>Pi20</i> (t)	<i>Pi20</i> (t)	<i>Pi20</i> (t)	ı		<i>Pi20</i> (t)		<i>Pi20</i> (t)	<i>Pi20</i> (t)	<i>k-m</i> , and
	C653-46	R I	М	К	К	Я	R I	R /	R /	К	R	R /	К	R I	R I	c-h, Pi
	B90002	К	R-M	Ч	К	Ч	К	Ч	Ч	ĸ	R	К	К	К	К	ik, Pil
	961058A	S	Ч	S	К	К	К	М	К	ĸ	R	Я	К	К	К	Pik; P
	607NB	S	S	R-M	Я	R	К	R	R	ы	R	R	R	Я	Я	les of
ate	2-12-2-1-6EM	К	\mathbf{v}	S	s	\mathbf{v}	К	Ч	Ч	\mathbf{v}	S	Я	s	К	R	le alle
Standard differential blast isolate	I-6-E-I-79W	S	Ч	\mathbf{v}	s	R	s	Х	К	К	S	s	К	ы	Ч	multip
ntial bla	1-8-5-1-65M	S	ч	\mathbf{v}	s	К	s	ч	Ч	ы	S	S	К	К	К	fy the
lifferer	952058Λ	S	ч	\mathbf{v}	s	\mathbf{v}	s	\mathbf{v}	\mathbf{v}	Μ	R	К	К	ы	Ы	classi
ndard o	C ^g 89	S	\mathbf{v}	\mathbf{v}	s	\mathbf{v}	s	\mathbf{v}	\mathbf{v}	\sim	R	К	К	s	\mathbf{v}	un not
Sta	IIIN	К	\mathbf{v}	\mathbf{v}	s	\mathbf{v}	К	R	R	\sim	R	R	К	К	К	ines ca
	E-18MI	S	Ч	\mathbf{v}	s	R-M	s	\mathbf{v}	\mathbf{v}	×	Μ	К	К	К	К	unt. hilipp
	9-9Od	S	\mathbf{v}	Ч	s	\mathbf{v}	s	\mathbf{v}	\mathbf{v}	S	R	Я	Ч	s	Ы	resista <i>Pik-s</i> 1 the P
	IK81-72	S	Ч	\mathbf{v}	s	R-M	s	\mathbf{v}	\mathbf{v}	К	\mathbf{S}	s	К	К	R	nd R; ect for s from
	1-01-E-1-9EM	К	\mathbf{v}	\mathbf{v}	s	\mathbf{N}	R	ч	Ч	\sim	Μ	К	S	К	R	tant, a ne exp isolate
	Monogenic lines and IRRI-bred varieties	IRBL20-IR24	IRBLta-CP1	IRBLk-K	IR20, IR28, IR30, IR45, IR66	IR29, IR34	IR8, IR22, IR24, IR26, PSBRc30	IR43	PSBRc2	IR5, IR32, IR36, IR38, IR40, IR42, IR44, IR50, IR52, IR54, IR58, IR60, IR62, IR65, IR68, IR72, PSBRc4	PSBRc1	IR74	IR56, IR70	IR46, IR48, IR64, PSBRc28	PSBRc10, PSBRc18, PSBRc20	S; susceptible, M; moderate resistant, and R; resistant. <i>Pik*</i> : One of the allele of <i>Pik</i> gene expect for <i>Pik-s</i> . These standard differential blast isolates from the Philippines can not classify the multiple alleles of <i>Pik</i> ; <i>Pik</i> , <i>Pik-h</i> , <i>Pik-m</i> , and <i>Pik-p</i> . <i>Pix</i> : Unknow gene
	Variety Group				VGla	VG1b	VG2a	VG2b	VG2c	VG3	VG4	VG5	VG6	VG7a	VG7b	S; suscel <i>Pik*</i> : On These ste <i>Pix</i> : Unk

Identification of blast resistance genes in IRRI-bred rice varieties by segregation analysis based on a differential system

Only one variety, PSBRc1 was resistant to PO6-6, susceptible to IK81-25, and moderately resistant to M36-1-3-10. This variety was classified as VG4 and estimated to harbor *Pik** but not *Pita* (Table 4). *Pi20*(t) was thought to be absent in PSBRc1, because this variety was susceptible to M39-1-2-21-2 which is incompatible with the gene (Table 2 and 5). The moderate resistance of this variety to M36-1-3-10 was probably conferred by unknown gene(s) other than genes in DVs tested.

IR74 was susceptible to only 1 isolate IK81-25 among the 3 isolate for primarily estimation of 3 genes. This variety was classified as VG5 and estimated to harbor Pi20(t) and Pik^* among the 3 genes (Table 4). The presence of these genes did not contradict to the susceptible reaction to M39-1-8-1 and M64-1-3-9-1 of this variety (Table 5).

Two varieties, IR56 and IR70 were resistant to PO6-6 and IK81-25 and susceptible to M36-1-3-10-1 (Table 4). These varieties were classified as VG6 and estimated to harbor *Pita* and *Pik** among the 3 selected genes for primary estimation. The reaction pattern of VG6 was corresponded with that of additional effect pattern of IRBLta-CP1 with *Pita* and IRBLk-Ka with *Pik* (Table 2 and 5).

Four varieties, IR46, IR48, IR64, and PSBRc28, were resistant to 2 isolates, M36-1-3-10-1 and IK81-25, and estimated to have Pi20(t) and Pita by primary estimation and classified as VG7 (Table 4). Three varieties, PSBRc10, PSBRc18, and PSBRc20 were resistant to PO6-6 in addition to those 2 isolates. These 3 varieties were estimated to have Pi20(t) and Pita but not Pik*, because these varieties were susceptible to Ca89 which was avirulent to Pik* (Table 5). These 3 varieties were classified as VG7b, while the former 4 were classified as VG7a. These 7 varieties were resistant to BN209 which was avirulent to Pik* and Pib (Table 2 and 5). Pik* was not considered to be harbored in these varieties as mentioned earlier. Therefore, Pib was estimated in these varieties in addition to Pi20(t) and Pita. Resistance to the blast isolate, PO6-6 in varieties classified as VG7b, was estimated to be due to the presence of Pii or Pi3.

II. Segregation analyses using backcross populations

Materials and methods

1. Rice plant materials

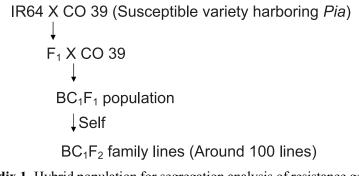
As shown in table 3, resistance to a certain blast isolate is governed by one or more genes in a variety. Segregation analysis using the BC_1F_2 family lines was conducted to determine the number of genes conferring resistance to the specific blast isolate. Six IRRI varieties representing several VGs, IR34 (VG1b), IR36 and IR60 (VG3), IR74 (VG5), and IR46 and IR64 (VG7a), were crossed with a susceptible check variety CO39 (a case of IR64 is shown in Fig. 1). The F_1 plants were backcrossed with CO39 as the recurrent parent to generate BC_1F_1 . The BC_1F_2 family lines with population sizes ranging from 53 to 104 were generated by selfing and used for the segregation analysis.

2. Inoculation and evaluation of resistance

The same procedure described above was employed for blast inoculation test. Differential blast isolates from the Philippines with known pathogenecity were used for the inoculation test to identify resistance genes. Each line consisting of 20 seedlings was inoculated with selected blast isolates avirulent to the expected genes.

3. Estimation of number of resistance genes

Following the procedure proposed by Toriyama et al. (1986), the number of genes conferring resistance in a variety was estimated from the segregation of BC_1F_2 lines to each blast isolate. Three ratios, 1:1, 3:1, and 7:1, for segregating for resistance (heterozygote) against susceptible (homozygote) lines were expected for one, two, and three genes controlling resistance, respectively (Fig. 2, Fig. 3, and Fig. 4, respectively). For example as shown in Fig. 2, where a single dominant gene in target rice



Appendix 1. Hybrid population for segregation analysis of resistance genes in the IRRI bred variety IR64

The hybrid population was developed by back crossing between IR64 and the susceptible variety CO 39 as the recurrent parent.

variety confers the resistance, ratio of number of resistant against susceptible BC_1F_1 plants and that of segregating against susceptible BC_1F_2 family lines are 1:1.

Segregation analysis using F_2 or F_3 family lines are also possible for estimation of number of resistance genes, although it was not employed in this study. The expected ratios for all resistant plants : segregation : all susceptible plants are 1:2:1 and 7:8:1 in F_3 family lines for a single and two dominant gene(s) control, respectively (Fig. 5 and 6, respectively).

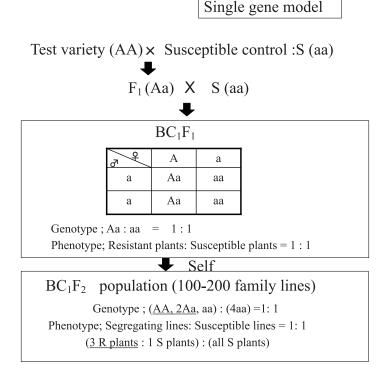
Results and discussion

1. Estimation of number of resistance genes in IR64

In IR64 of VG7a, 72 BC₁F₂ lines were generated by backcross with CO39 (Fig. 1). These lines were tested against four isolates, BN111, IK81-25, M64-1-3-9-1, and BN209 (Table 6). The ratios of number of all resistant lines (resistant homozygote) : segregating lines (heterozygote) : all susceptible lines (susceptible homozygote) in response to two isolates, BN111 and IK81-25 were well fit to 0:1:1 (χ^2 =0.06, *p*=0.81, and χ^2 =1.39, *p*=0.24, respectively). These results suggested that the resistance to each of these 2 isolates was controlled by a

single dominant gene. The ratios in number of lines of all resistant : segregating : all susceptible to the remaining two isolates, M64-1-3-9-1 and BN209 were well fit to 0:3:1 (χ^2 =0.67, *p*=0.41 and χ^2 =0.08, *p*=0.78, respectively). From these results, it was estimated that the resistance to each of these 2 isolates was controlled by 2 dominant genes.

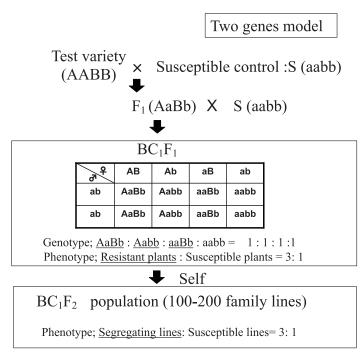
Isolate BN111 is avirulent to Pi20(t) and Pik^* (Table 3). A single dominant gene controlling resistance of IR64 to this isolate was estimated to be Pi20(t) but not Pik^* , because the gene Pik^* was not estimated in IR64 in the previous estimation by differential system (Table 5 and 6). Similarly, a single dominant gene for resistance to IK81-25 was estimated to be Pita (Table 3 and 6). The remaining 2 blast isolates, M64-1-3-9-1 and BN209 which were estimated to be controlled by 2 genes, were avilurent to Pita and Piz-t, and to Pib, respectively (Table 3). From these results, four genes (Pi20(t), Pita, Piz-t, and Pib) were detected in IR64 to control the segregation for resistance to the isolates used. However, one of the two genes could not be identified in the segregation analysis using BN209.



Appendix 2. Segregation analysis for blast resistance gene using BC₁F₂ plants or BC₁F₂ family lines – single gene model

A; resistance gene harbored in the genetic background. Where a single dominant gene in a test rice variety confers resistance, the ratio of number of resistant to susceptible BC_1F_1 plants and that of segregating to susceptible BC_1F_2 family lines is 1 : 1.

Avirulence blast pathogen for the resistance gene, A, is used in segregation analysis. Avirulence pathogen is selected by prior inoculation test on the test variety. When the virulent blast isolate is used, all progeny will be susceptible.

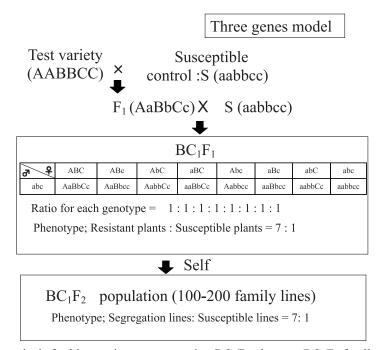


Appendix 3. Segregation analysis for blast resistance gene using BC_1F_2 plants or BC_1F_2 family lines – two gene model

A, B; Resistance genes harbored in the genetic background.

Where two dominant genes of test rice variety confer resistance independently, the segregation ratio in the BC_1F_1 plants for resistant (heterozygote) and susceptible (homozygote) plants is 3:1. Accordingly, the ratio of number of BC_1F_2 family lines segregating for resistance to all susceptible lines is 3:1.

A blast isolate avirulent to the differential variety with a known resistance gene is used for the inoculation test.

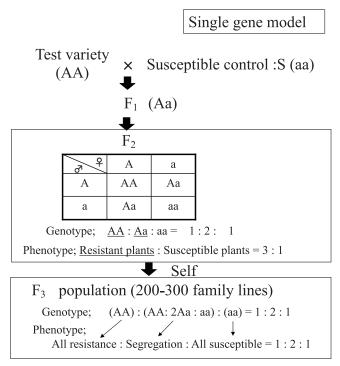


Appendix 4. Segregation analysis for blast resistance gene using BC₁F₂ plants or BC₁F₂ family lines – three gene model

A, B, C; Resistance genes harbored in the background.

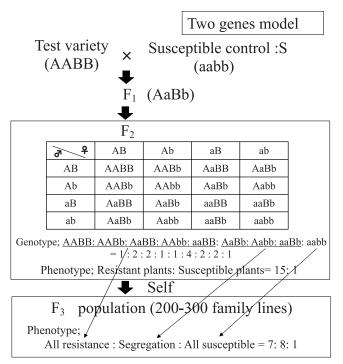
Where three dominant genes in the genetic background of test rice variety confer resistance independently, the segregation ratio in the BC_1F_1 plants for resistant (heterozygote) and susceptible (homozygote) plants is 7:1. Accordingly, the ratio of number of BC_1F_2 family lines with segregating for resistance to all susceptible lines is 7:1.

A blast isolate avirulent to the differential variety with known resistance gene is used for the inoculation test.



Appendix 5. Segregation analysis for blast resistance genes using F_2 plants or F_3 family lines – single gene model

A; Resistance gene harbored in the genetic background. Where a single dominant gene in the genetic background of a test rice variety confers resistance, the ratio for the number of resistant homozygote : heterozygote : susceptible homozygote F_2 plants is 1:2:1. Accordingly, the ratio for number of F_3 family lines is 1(all R): 2 (segregation): 1 (all S). A blast isolate avirulent to the differential variety with the known resistance gene is used for the inoculation test.



Appendix 6. Segregation analysis for a blast resistance gene using F2 plants or F3 family lines - two gene model

A, B; Resistance genes harbored in the genetic background. Where two dominant genes in the genetic background of a test rice variety confer resistance independently, the ratio for the number of resistant homozygotes with at least one gene: heterozygote : susceptible homozygote F_2 plants is 7:8:1. Accordingly, the ratio for the number of F_3 family lines is 7 (all R): 8 (segregation) : 1 (all S). A blast isolate avirulent to the differential variety with the known resistance gene is used for the inoculation test.

2. Estimation of number of resistance genes in IRRIbred varieties

Table 7 shows the results of the segregation analyses in six BC₁F₂ populations. In IR34 (VG1b), three genes, Pik-s, Pib, and Piz-t, were previously estimated (Table 5), hence, genetic analysis was performed to confirm these genes. A total of 104 BC_1F_2 family lines were examined for segregation to five isolates, M39-1-3-8-1, M64-1-3-9-1, V850196, BN209, and B90002. Against M39-1-3-8-1 and M64-1-3-9-1, 48 lines were segregating resistant and 56 lines were susceptible homozygote, and a co-segregation for resistance was recognized between these isolates. The segregation fit a 1:1 ratio expected of a single dominant gene control. Since both isolates were avirulent to Piz-t and Pita, but IR34 in VG1 was previously estimated not to have Pita, the segregation for resistance was therefore governed by *Piz-t*. To isolates V850196 and BN209, the segregation rates between segregating resistant and susceptible homozygote lines fit a 3:1 ratio, suggesting that two dominant genes controlled the resistance to each isolate. One of the genes was identified as Pik-s to V850196 based on the estimation by reaction patterns, but the other one could not be identified by the differential system. Similarly, *Pib* was identified as one of the resistance genes to BN209 but the other one was not identified. The same lines were also examined for Pia, a resistance gene that was not estimated by reaction patterns because Pia was masked by *Pi20*(t), *Pita*, or *Pik**. All the lines showed a resistant reaction to isolate B90002, which was avirulent to Pia. Therefore, Pia was identified in IR34 since it was known to be included in the recurrent parent, CO39 (Tsunematsu et al., 2000). On the basis of these results, the presence of Pib, Pik-s, and Piz-t was confirmed in IR34, while another gene Pia and at least an unknown one were also detected.

Two varieties from VG 3, IR36 and IR60, previously estimated to harbor *Pita* and *Pib*, were analyzed (Table

7). Results showed that Pita could be identified in IR36 using isolates IK81-3, and V850196 which are avirulent to *Pita*. The 61 BC_1F_2 lines derived from a cross between IR36 and CO39 exhibited single gene segregation to IK81-3. The segregation for resistance was due to Pita. To isolates V850196, the segregation rates between segregating resistant and susceptible homozygote lines fit a 3:1 ratio, suggesting that two dominant genes controlled the resistance. Therefore, Pib, which was previously estimated and incompatible to this isolate, was identified in IR36 as one of the gene conferring the resistant to this isolate. Pik-s was also identified in IR36 as another resistance gene to V850196 with considering the reaction pattern (Table 5). Similarly, the resistance to BN209 that has avirulence to Pib and Pik* among the gene tested, was considered to be controlled by 2 dominant genes. Previous analysis showed that Pik* was not included in VG 3; hence, segregation for resistance to BN209 was related to Pib and another unknown gene. As a result of segregation analysis, 3 genes, Pib, Pita and Pik-s as well as one unknown gene were confirmed to be harbored in IR36.

The presence of two genes, *Pita* and *Pib*, was analyzed in IR60 of VG3. Using 53 BC₁F₂ lines, a digenic segregation to each of the three isolates, IK81-3, M64-1-3-9-1, and V850196 was obtained, and three genes controlled were estimated for resistance to BN209 (Table 7). IK81-3 is avirulent to *Pita* and *Pik**, while the latter was not present in VG3 by the previous estimation. M64-1-3-9-1 was avirulent to *Pita* and *Piz-t*, V850196 to *Pita* and *Pik-s*, and BN209 to *Pib* and *Pik** in the differential system. Thus, 3 genes, *Pita*, *Pik-s*, and *Pib*, were considered to be related in the segregation. However, the other one gene for resistance to IK81-3 and two genes for resistance to BN209 could not be identified by reaction patterns of the differential system.

IR74 in VG5 was estimated to harbor 2 genes, Pi20(t) and Pik^* by reaction patterns (Table 5). This variety was

	Read		f monog ine	genic		Nc	o. of BO	C ₁ F ₂ li	nes		χ^2 v	alue			
Differntial blast isolate	IRBL20-IR24 (<i>Pi20</i>)	IRBLta-K1(Pita)	IRBLb-B (Pib)	(Pib) (Piz-t)		All resistant plants	Segregating	All susceptible	Total	-	1:1	3:1	P (df =1)	Estimated no. of resistance gene	Estimated gene
BN111	R	S	S	S		0	37	35	72		0.06	-	0.81	1	<i>Pi20</i> (t)
IK81-25	S	R	S	S		0	41	31	72		1.39	-	0.24	1	Pita
M64-1-3-9-1	S	R	S	R		0	51	21	72		-	0.67	0.41	2	Pita, Piz-t
BN209	S	S	R-M	S		0	52	17	69		-	0.08	0.78	2	Pib, one unknown

Table 6. Segregation for resistance in BC_1F_2 populations derived from a cross between IR64 and a susceptible variety CO 39 as the recurrent parent

analyzed using 76 BC₁F₂ lines for segregation to six isolates: PO6-6 and Ca89 for identification of *Pik**, M36-1-3-10-1 for *Pi20*(t), BN111 for *Pi20*(t) and *Pik**, BN209 for *Pib* and *Pik**, and B90002 for *Pia*, *Pik**, *Pi20*(t), *Pib*, and *Piz-t* (Table 7). In the segregation analysis on BC₁F₂ lines, resistance was controlled by a single dominant gene judging from the segregation ratio of 1:1 involving three isolates, PO6-6, Ca89, and M36-1-3-10-1. A cosegregation was also observed between isolates PO6-6 and Ca89. *Pik** was identified to control the segregation for PO6-6 and Ca89, and *Pi20*(t) was for isolate M36-1-3-10-1. A digenic segregation to isolates BN111 and BN209 were observed. One of the genes, *Pik**, was identified to be common in the segregation to both isolates.

Table 7. Segregation for resistance in BC_1F_2 populations derived from the crosses between IRRI-bredvarieties and a susceptible variety CO 39 as the recurrent parent

Variata			No. of	lines		χ	2 ² value	1)	- P	Estimated	Estimated
Variety (VG)	Isolate	All resistant	Segregating resistant	All susceptible	Total	1:1	3:1	7:1		no. of resistance gene	gene ²⁾
	M39-1-3-8-1	-	48	56	104	0.62 ^a	-	-	0.43	1	Piz-t
	M64-1-3-9-1	-	48	56	104	0.62 ^a	-	-	0.43	1	Piz-t
IR34 (VG1b)	V850196	-	77	27	104	-	0.02	-	0.89	2	<i>Pik-s</i> , one unknown
(()010)	BN209	-	79	25	104	-	0.02	-	0.89	2	<i>Pib</i> , one unknown
	B90002	104	0	0	104	-	-	-	-		Pia
	IK81-3	-	35	26	61	1.32	-	-	0.25	1	Pita
IR36	V850196	-	47	14	61	-	0.13	-	0.72	2	Pita, Pik-s
(VG3)	BN209	-	45	16	61	-	0.05	-	0.82	2	<i>Pib</i> , one unknown
	IK81-3	-	36	17	53	-	1.41	-	0.24	2	<i>Pita</i> , one uknown
IR60	M64-1-3-9-1	-	40	13	53	-	0.02	-	0.89	2	Pita, Piz-t
(VG3)	V850196	-	37	16	53	-	0.76	-	0.38	2	Pita, Pik-s
	BN209	-	48	5	53	-	-	0.46	0.5	3	<i>Pib</i> , two unknown
	M36-1-3-10-1	-	36	40	76	0.22	-	-	0.64	1	<i>Pi20</i> (t)
	BN111	-	51	25	76	-	2.52	-	0.11	2	<i>Pi20</i> (t), <i>Pik</i> ^{*3)}
IR74	PO6-6	-	34	42	76	0.84 ^b	-	-	0.36	1	Pik*
(VG5)	Ca89	-	34	42	76	0.84 ^b	-	-	0.36	1	Pik*
	BN209	-	54	22	76	-	0.63	-	0.43	2	Pik*, Pib
	B90002	76	0	0	76	-	-	-	-		Pia
	M36-1-3-10-1	-	58	42	100	2.56 ^d	-	-	0.11	1	<i>Pi20</i> (t)
	BN111	-	58	42	100	2.56 ^d	-	-	0.11	1	<i>Pi20</i> (t)
ID 47	IK81-25	-	59	41	100	3.24 ^c	-	-	0.07	1	Pita
IR46 (VG7a)	M64-1-3-9-1	-	59	41	100	3.24 ^c	-	-	0.07	1	Pita
,	V850196	-	79	21	100	-	0.85	-	0.36	2	Pita, Pik-s
	BN209	-	81	19	100	-	1.92	-	0.17	2	<i>Pib</i> , one unknown
	BN111	-	37	35	72	0.06	-	-	0.81	1	<i>Pi20</i> (t)
IR64	IK81-25	-	41	31	72	1.39	-	-	0.24	1	Pita
(VG7a)	M64-1-39-1	-	51	21	72	-	0.67	-	0.41	2	Pita, Piz-t
	BN209	-	52	17	69	-	0.08	-	0.78	2	<i>Pib</i> , one unknown

1) Common letters (a, b, c, and d) indicate co-segregation among isolates.

2) Resistance genes described by bold letters were those estimated by the differential system (see Table 5).

3) Pik*: One of the alleles Pik, Pik-h, Pik-m, or Pik-p.

The other genes segregating to BN111 and BN209 corresponded to *Pi20*(t) and *Pib*, respectively. In the analysis of segregation to B90002, all the lines were resistant, suggesting that IR74 carries the same *Pia* gene found in CO39. These results revealed that besides *Pita* and *Pik** confirmed in IR74, two other genes, *Pia* and *Pib* were identified in this variety.

Two varieties in VG 7a, IR46 and IR64, were analyzed to confirm the presence of Pi20(t), Pita, and Pib. In the analysis of IR46 involving 100 BC₁F₂ lines and four isolates, M36-1-3-10-1, BN111, IK81-25, and M64-1-3-9-1, a 1:1 ratio suggesting the monogenic segregation, and the co-segregation for resistance to BN111 and M36-1-3-10-1, and to IK81-25 and M64-1-3-9-1 were observed (Table 7). Since BN111 and M36-1-3-10-1 are avirulent to Pi20(t), and IK81-25 and M64-1-3-9-1 to Pita, the presence of Pi20(t)and Pita in IR46 was confirmed. A 3:1 ratio observed between segregating resistant and susceptible homozygote lines suggested that the segregation for resistance to isolates V850196 and BN209 was controlled by two dominant genes. In the reaction pattern of differential system, V850196 was avirulent to Pita, Pik* and Pik-s, and BN209 to Pib and Pik*. The gene Pik* was estimated not to be harbored in VG 7a by reaction pattens. The other genes, Pita and Pik-s, were related to the segregation for resistance to V850196, and Pib and an unknown gene to BN209. Thus, Pi20(t), Pita, Pik-s, and Pib, and one unknown gene were identified in IR46.

III. Allelism tests of the resistance genes using differential varieties

Materials and methods

1. Rice plant materials

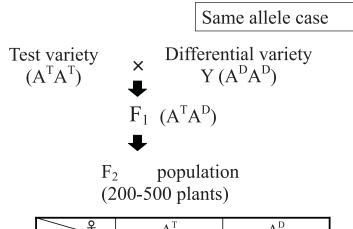
Nine IRRI rice varieties from 7 different VGs, IR34 (VG1b), IR24 (VG2a), IR36 and IR60 (VG3), PSBRc1 (VG4), IR74 (VG5), IR56 and IR70 (VG6), and IR64 (VG7b), were crossed with DVs with known blast resistance genes. The DVs were Toride 1 for *Piz-t*; BL 1 for *Pib*; Yashiromochi, C105TTP2L9, and C101PKT for *Pita*; Tsuyuake with *Pik-m* and Kanto 51 with *Pik* for *Pik**; and IR24 for three genes, *Pib*, *Pik-s*, and *Pi20*(t) (Imbe et al.1997). The F₂ populations derived from each cross were tested.

2. Inoculation and evaluation of resistance

The same procedure described above was employed for blast inoculation test. The F_2 populations were inoculated with selected blast isolates, which were avirulent to particular genes.

3. Identification of resistance gene

The allelic relationship of the genes was determined based on the segregation for resistance to a particular blast isolate in the F_2 populations. If the segregation did not include susceptible plants, it was assumed that both parents had the same or an allelic gene (Fig. 7). Otherwise, it was considered that the resistance was governed by different allele genes (Fig. 8).



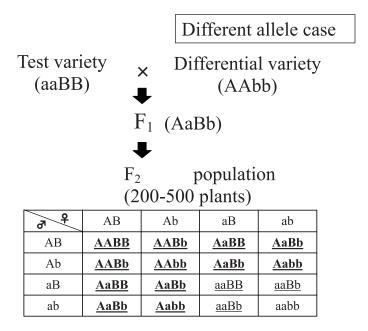
of 7	A^{T}	A^{D}
A^{T}	$A^{T}A^{T}$	$A^{D}A^{T}$
A ^D	$A^{T}A^{D}$	A ^D A ^D

Phenotype; All resistant plants

Appendix 7. Allelism test using an F₂ population derived from a cross between a test rice variety and differential varieties – a case of the same allele

AT, AD; resistance gene in the rice test variety and differential variety, respectively. Inoculation test is carried out with a blast isolate avirulent to the differential variety.

Identification of blast resistance genes in IRRI-bred rice varieties by segregation analysis based on a differential system



•Segregation ratio in case of inoculation with a blast isolate avirulent only to the differential variety.

 \rightarrow Resistance plants: Susceptible plants = 3 : 1

•Segregation ratio in case of inoculation with a blast isolate avirulent

to both the differential and the test variety.

 \rightarrow <u>Resistance plants</u> : Susceptible plants = 15 : 1

Appendix 8. Allelism test using F₂ population derived from the cross between rice test variety and differential variety – a case of different allele

A, B; resistance gene in the differential and the test variety, respectively.

A blast isolate avirulent to the differential variety carrying the known resistance gene is used for the inoculation test.

Results and discussion

1. Gene identification in IR64

The F_2 populations were generated from crosses between IR64 of VG7a and 5 DVs, IR24 for Pi20(t), Piks and Pib, C105TTP2L9 and C101PKT for Pita, Toride 1 for *Piz-t*, and IR56 for *Pita* and *Pik**. When the F₂ populations derived from cross between IR64 and IR24 were tested against 3 blast isolates, BN111 avirulent to Pi20(t) and Pik, V850196 avirulent to Pita, Pik and Pik-s, and BN209 avirulent to *Pik* and *Pib*, no population shows any susceptible plants to these isolates (Table 8). These results indicated that IR64 was revealed to harbor the same alleles of Pi20(t), Pik-s and Pib in IR24 by inoculation test against BN111, V850196 and BN209, respectively. The F₂ plants derived from crosses between IR64 and 2 DVs for Pita, C105TTP2L9 and C101PKT were all resistant to a blast isolate, IK81-25 avirulent to *Pita*. This suggested that IR64 also harbored the same allele of Pita in those DVs. Similarly, Piz-t was identified in IR64, when the F₂ plants derived from cross between IR64 and Toride 1 which is a DV for Piz-t were all resistant to both M39-1-3-8-1 and M64-1-3-9-1 which are avirulent to the gene. In the F_2 population derived from a cross between IR64 and IR56, all tested plants were

resistant to IK81-25 avirulent to *Pita*. On the other hand, a 3:1 segregation ratio ($\chi^2 = 0.22$, p=0.64) was observed in the same population for resistance to PO6-6, which is avirulent to *Pik**. This resistance was suggested to be governed by a single gene, *Pik** in IR56. These results confirmed that IR64 harbored 6 resistance genes, *Pi20*(t), *Pita*, *Pib*, *Pik-s*, *Piz-t*, and one unknown.

2. Gene identification in IRRI-bred varieties

To confirm the presence of Piz-t and Pib in IR34 in VG1b, F₂ populations derived from crosses with 2 DVs, Toride 1 for Piz-t and BL 1 for Pib were tested against the selected blast isolates. The F₂ population derived from cross with Toride 1 was inoculated with 2 blast isolates, M64-1-3-9-1 and M39-1-3-8-1 both avirulent to Piz-t (Table 9). As a result, all plant of the population was resistant to these isolate. Thus, the resistance gene in IR34 to these isolate was the same as that in Toride 1, *Piz-t*. In the same manner, F₂ population derived from cross with BL1 did not show any susceptible plants to blast isolate BN209 avirulent to *Pib*. These results suggested the presence of *Piz-t* and *Pib* in IR34.

All F₂ plants derived from the cross between IR24 in VG2a and a DV, BL 1 for *Pib* was resistant to BN209

avirulent to *Pik** and *Pib* (Table 9). These results suggested the presence of *Pib* in IR24 and supported the previous report by Imbe et al. (1997).

Four kind of F₂ population were generated from crosses between IR36 in VG3 and 4 DVs; C105TTP2L9 and C101PKT for Pita, Toride 1 for Piz-t, and BL 1 for *Pib.* When these populations were tested against 4 isolates, IK81-25 (avirulent to Pita), M391-3-8-1 and M64-4-1-3-9-1 (avirulent to Pita and Piz-t), and BN209 (avirulent to Pib), all F₂ plants showed resistance. These results confirmed the presence of Pita, Piz-t, and Pib in IR36. The presence of *Piz-t* was also confirmed by the allelism test between IR34 and IR36 using M39-1-3-8-1 and M64-1-3-9-1. Using F₂ population derived from a cross between IR36 and IR24, the allelism of Pib and Pik-s was confirmed by inoculation test against blast isolate BN209 (avirulent to Pib) and V850196 (avirulent to *Pik-s*), respectively. The reaction of *Piz-t* was masked by *Pita* and could not be estimated in the previous analysis based on reaction pattern. However, Piz-t was identified in IR36 through allelism tests. Thus, BC1F2 analysis and allelism tests revealed that IR36 carries at least four genes-Pita, Pik-s, Pib, and Piz-t.

The F_2 populations derived from crosses of IR60 in VG3 with IR34 and IR24 did not segregate any susceptible plants to 4 isolates, BN209 (avirulent to *Pib* and

*Pik**), V850196 (avirulent to *Pita*, *Pik-s* and *Pik**), M39-1-3-8-1 and M64-1-3-9-1 (avilurent to *Pita* and *Piz-t*) (Table 9). IR34 was confirmed to harbor *Pia*, *Pib*, *Piz-t*, and *Pik-s* in this study. IR24 was also confirmed to harbor *Pia*, *Pib*, *Pik-s*, and *Pi20*(t), previously. IR60 was estimated not to harbor *Pik** by reaction pattern, while the resistance to M39-1-3-8-1 and M64-1-3-9-1 was governed by *Piz-t* which was estimated also in IR34. From these results, IR60 was revealed to have 3 genes, *Pib*, *Piz-t*, and *Pik-s*.

PSBRc1 of VG4 was crossed with IR56, which was estimated to be carrying *Pita* and *Pik** to generate F_2 population. All of the F_2 plants was resistant to isolates PO6-6 avirulent to *Pik** and BN111 avirulent to both *Pik** and *Pi20*(t) (Table 9). Since *Pi20* was not estimated in VG4 by reaction patterns in the previous analysis, the resistance of F_2 plants to these isolates suggested the allelism of *Pik** in PSBRc1 and IR56. The presence of *Pita* was also verified in this population using IK81-25 which is avirulent to *Pita*. Results showed that the F_2 plants segregated into 406 resistant and 126 susceptible, which fit the 3:1 ratio (χ^2 =0.51, p=0.43). This indicated that a single dominant gene, *Pita* previously estimated in IR56 governed the segregation, because PSBRc1 was estimated not to have *Pita* by reaction pattern.

Table 8. Allelism test for resistance using F₂ populations derived from the crosses between IR64 and differential varieties

		Differntial	R	eactio	n of r	nonoge	enic li	ne	No.	of pla	ants ¹⁾			
(Resistance g	ombination gene harbored in arent)	blast isolate used for the seregation analysis of resistance	IRBL20-IR24 (<i>Pi20</i>)	IRBLta-K1 (Pita)	IRBLk-Ka (<i>Pik</i>)	IRBLb-B (Pib)	IRBLzt-T (Piz-t)	IRBLks-F5 (Pik-s)	R	S	Total	χ^2 value (3:1)	P (df=1)	Identified) gene
IR24	IR64	BN111	R	S	R	S	S	S	302	0	302	-	-	Pi20(t)
(<i>Pib</i> , <i>Pi20</i> (t),	/ (Pi20(t), Pita,	V850196	S	R	R	S	S	R	227	0	227	-	-	Pik-s
Pik-s)	Pib)	BN209	S	S	R	R-M	S	S	222	0	222	-	-	Pib
IR64	/ C105TTP2L9 (<i>Pita</i>)	IK81-25	S	R	S	S	S	S	274	0	274	-	-	Pita
IR64	/ C101PKT / (Pita)	IK81-25	S	R	S	S	S	S	132	0	132	-	-	Pita
IR56 (Pita,	/ IR64	IK81-25	S	R	S	S	S	S	296	0	296	-	-	Pita
Pik*)	/ 1004	PO6-6	S	S	R	S	S	S	193	60	253	0.22	0.64	Pik* in IR56
Toride1	/ IR64	M39-1-3-8-1	S	R	S	S	R	S	124	0	124	-	-	Piz-t
(Piz-t)	/ 1604	M64-1-3-9-1	S	R	S	S	R	S	133	0	133	-	-	Piz-t

1) R; resistant, S; susceptible.

2) Pik*: One of the alleles Pik, Pik-h, Pik-m, or Pik-p.

IR24 was expected not to habor Pita based on the reaction pattern to differential blast isolates in previous reasearch. IR24 and IR64 were expected not to harbor Pik based on the reaction patterns to differential blast isolates in previous reasearch.

Toride 1 dose not harbor Pita

Variety		ss combination	Isolate	No	. of plar	nts ¹⁾	χ^2 value	<i>P</i>	Identified
(VG)	(Resistance ge	ene(s) harbored in parent)		R	S	Total	(3:1)	(df = 1)	gene
IR34	Toride1 (<i>Piz-t</i>)	/ IR34 (Pia, Pik-s, Pib, Piz-t)	M39-1-3-8-1	94	0	94	-	-	Piz-t
(VG1b)			M64-1-3-9-1	84	0	84	-	-	Piz-t
	BL1 (Pib)	/ IR34	BN209	437	0	437	-	-	Pib
IR24 (VG2b)	BL1	/ IR24	BN209	161	0	161	-	-	Pib
IR36	IR36 (Pita, Pib, Pik-s)	/ C105TTP2L9 (Pita)	IK81-25	468	0	468	-	-	Pita
(VG3)	IR36	/ C101PKT (Pita)	IK81-25	167	0	167	-	-	Pita
	BL1	/ IR36	BN209	420	0	420	-	-	Pib
	IR24	/ IR36	V850196	295	0	295	-	-	Pik-s
			BN209	287	0	287	-	-	Pib
	Toride 1	/ IR36	M39-1-3-8-1	184	0	184	-	-	Piz-t
			M64-1-3-9-1	168	0	168	-	-	Piz-t
	IR34	/ IR36	M39-1-3-8-1	467	0	467	-	-	Piz-t
			M64-1-3-9-1	516	0	516	-	-	Piz-t
IR60	IR34	/ IR60 (Pita, Pib, Pik-s, Piz-t)	M39-1-3-8-1	302	0	302	-	-	Piz-t
(VG3)			M64-1-3-9-1	303	0	303	-	-	Piz-t
	IR24	/ IR60	V850196	235	0	235	-	-	Pik-s
			BN209	360	0	360	-	-	Pib
PSBRc1	IR56 (Pita, Pik*)	/ PSBRc1 (<i>Pik</i> * ²)	PO6-6	531	0	531	-	-	Pik*
(VG4)			BN111	526	0	526	-	-	Pik*
			IK81-25	406	126	532	0.51	0.43	Pita in IR56
IR74	IR56	/ IR74 (Pi20(t), Pik*)	PO6-6	560	0	560	-	-	Pik*
(VG5)			BN111	585	0	585	-	-	Pik*
			IK81-25	442	135	577	0.82	0.37	Pita in IR56
IR56	C101PKT	/ IR56	IK81-25	469	0	469	-	-	Pita
(VG6)			PO6-6	358	85	443	7.98	0.005	Pik* in IR56
	Yashiromochi (Pita)	/ IR56	IK81-25	463	0	463	-	-	Pita
			PO6-6	370	127	497	0.08	0.78	Pik* in IR56
	IR56	/ Kanto 51 (Pik)	PO6-6	440	0	440	-	-	Pik*
			BN111	407	0	407	-	-	Pik*
	IR56	/ Tsuyuake (Pik-m)	PO6-6	498	0	498	-	-	Pik*
			BN111	496	0	496	-	-	Pik*
IR70	IR56	/ IR70 (Pita, Pik*)	IK81-25	567	0	567	-	-	Pita
(VG6)			PO6-6	577	0	577	-	-	Pik*
			BN111	558	0	558	-	-	Pik*

Table 9. Allelism test for resistance	using F ₂ populations	derived from the	crosses between	IRRI-bred and
differential varieties				

1) R; resistant, S; susceptible.

2) *Pik**; One of the alleles *Pik*, *Pik-h*, *Pik-m*, or *Pik-p*.

*Pik** was identified in IR74 by an allelism test with IR56 (Table 9). All the F₂ plants derived from a cross of IR74 with IR56 were resistant to PO6-6, which is avirulent to *Pik**, and BN111 avirulent to both *Pik** and *Pi20*(t), suggesting that IR74 had the same allele of *Pik** in IR56. The segregation ratio for resistance to IK81-25, which was avirulent to *Pita* fitted to a 3:1 ratio (χ^2 = 0.82, p=0.37) expected for a single dominant gene control. This was thought to be due to *Pita* in IR56 but not in IR74.

In VG6, two genes, *Pita* and *Pik**, were previously estimated by reaction patterns in IR56 and IR70. To confirm their presence in these varieties, IR56 was crossed with 4 DVs, C101PKT and Yashiromochi for Pita, Kanto 51 and Tsuyuake for Pik*, and with IR70 for Pita and Pik*. On these 5 kinds of F2 populations, 11 combinations of allelism test were carried out using 3 isolates: IK81-25 avirulent to Pita, PO6-6 to Pik*, and BN111 to *Pik*^{*} and *Pi20*(t). Of these, two F_2 populations derived from crosses of IR56 with C101PKT and Yashiromochi showed a single gene segregation to PO6-6, suggesting that the segregation was due to Pik^* in IR56 (Table 9). The other 9 combinations did not segregate any susceptible plants, suggesting that the same gene in IR56 or IR70 and DVs controlled the resistance. Since blast isolates from the Philippines could not differentiate the Pik alleles, except Pik-s, no susceptible plants were obtained from the populations derived from the combinations among DVs, Kanto 51, Tsuyuake, and IR56 for Pik*. These results confirmed that IR56 and IR70 harbored Pita and Pik*.

Conclusions

A total of 42 IRRI-bred rice varieties were classified into seven variety groups and estimated at least seven blast resistance genes, Pi20(t), Pita, Pik*, Pib, Pik-s, Piz-t, and Pii or Pi3(t), following a differential system using Philippine isolates of P. grisea (Ebron et al., 2004, Fukuta et al. 2007). In the present study, the presence of seven blast resistance genes, Pi20(t), Pita, Pik*, Pia, Pib, Pik-s, and Piz-t, was confirmed through genetic analyses in 10 varieties: IR34 (VG1b), IR24 (VG2a), IR36 and IR60 (VG3), PSBRc1 (VG4), IR74 (VG5), IR56 and IR70 (VG6), and IR46 and IR64 (VG7a) (Table 5, 7, and 9). Four of these genes, Pia, Pib, Pik-s, and Piz-t, were identified also in some varieties. Two genes Pib and Pik-s or Pik* were previously estimated in nearly all the varieties and their presence confirmed in the present study. The genes Pita, Pik alleles (Pik* and *Pik-s*), *Piz-t*, and *Pib* were identified frequently in Indica-type varieties (Kiyosawa, 1966; Kiyosawa and Murty, 1969; Yokoo and Kiyosawa, 1970; Yokoo et al., 1978). Our results agreed with the findings of these previous studies.

The present study reports the genotypes of blast resistance in IRRI-bred rice varieties determined by segregation analyses using backcrossed progenies and allelism tests with DVs to confirm the estimated resistance genes and to postulate the masked ones in the IRRI-bred varieties' genetic backgrounds. The utility of genetic analyses based on the differential system was demonstrated in this study, and this information will be very useful whenever IRRI-bred varieties are used in any rice breeding program. Genetic analyses based on the differential system are useful tools for the identification of resistance genes, although these are complicated and relatively tedious. Over the past several years, two genes Pita and Pib have already been isolated and their detailed sequences described by Wang et al. (1999) and Jia et al. (2002), respectively. Based on this information, molecular markers specific to both genes were developed and can be used to identify the genotypes of Pib and Pita in Indicatype varieties. Therefore, Indica-type, IRRI-bred varieties, which harbor several resistance genes and show complex reactions, can be determined of these genotypes using molecular markers that can complement detection of the genes by standard genetic analysis.

The differential system, which is based on conventional methods and which does not require advanced facilities, was revealed to be useful as a fundamental tool to provide essential information to develop breeding programs for blast resistance.

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