Simple and Rapid DNA Extraction from Milled Rice and Its Application to Thai Aromatic Rice (*Oryza sativa* L.) variety, Khao Dawk Mali 105

Tadashi YOSHIHASHI

Crop Production and Postharvest Technology Division
Japan International Research Center for Agricultural Sciences (JIRCAS)
(Tsukuba, Ibaraki, 305-8686 JAPAN)

Received September 21, 1999

Abstract

An aromatic variety, Khao Dawk Mali 105 (KDML 105), is one of the most important varieties of rice in Thailand. However, the increase of production is limited by adverse environmental conditions. As a result, varieties different from KDML 105 are often marketed under the name of KDML 105. Therefore, an efficient and objective method of cultivar identification is urgently needed. Molecular marker techniques based on the polymerase chain reaction (PCR), have provided objective methods of identification. To apply this technique to milled rice, high quality DNA from a single grain of milled rice is needed. DNA extraction method presented here is based on a freeze-and-thaw cycle that enables to extract purified high molecular weight grain DNA without the use of expensive equipment and/or time-consuming procedures. Microsatellite analysis method for cultivar identification was used for extracted DNA. The author analyzed rice samples obtained from Bangkok, Suwannaphum, and Kuala Lumpur markets, and selected varieties different from KDML 105 in Bangkok and Kuala Lumpur market rice samples. By this extraction procedure, we were able to obtain DNA for microsatellite analysis within 2hr and identification required 6hr in total.

Key words: cultivar identification, PCR, rapid extraction, genomic DNA
Introduction

A large quantity of rice is produced in the world presently. Rice farmers need to produce rice which can meet the demand of consumers. Aromatic rice, which has a stronger aroma than ordinary rice, has been very popular in Southeast Asia\(^5\)\(^6\) and has recently gained a wide acceptance in Europe and U.S.A. An aromatic variety, Khao Dawk Mali 105 (KDML105), is mainly produced in Northeast Thailand. Demand for this variety is increasing in both domestic and international markets, due to the recognition of its good quality. Although increase of production is urgently needed, the cultivation is limited due to infertile and drought-stricken sandy soil. Erratic rainfall at the beginning of the rainy season and labor shortage for transplanting are other constraints on the production of KDML 105. As a result, varieties different from KDML 105 are often marketed under the name of KDML 105, because no objective method of evaluation has been developed for this variety.

In Japan, since rice is distributed as brown rice which can germinate, information about the varieties can be obtained. However, in Southeast Asia, rice is mainly distributed as milled rice which cannot germinate. To identify varieties within milled rice mixed with varieties with a similar appearance, conventional methods like the determination of the length-width ratio are not suitable.

Molecular marker techniques based on the polymerase chain reaction (PCR)\(^10\), have provided more objective methods for the identification of grain varieties. These techniques like microsatellite analysis, randomly amplified polymorphic DNA (RAPD)\(^12\) and amplified fragment length polymorphism (AFLP)\(^7\) could be very useful for plant-based identification.\(^4\)\(^14\) Microsatellite analysis is particularly useful for simple and rapid analysis. To apply this technique to milled rice, high quality DNA must be extracted from a single grain of milled rice. Preparation of DNA from single rice seedlings was reported\(^11\), but milled rice cannot germinate.

Components of milled rice are mainly starch and protein. It is difficult to extract DNA, because the solubility of these components is the same as that of DNA. Especially, enzyme-inhibiting polysaccharides are often present in the “purified” DNA. Extraction procedures for simple and rapid identification of grain variety require that cell walls be broken, cell membrane be disrupted, DNA be protected from endogenous nucleases, and polysaccharides be removed. Murray & Thompson\(^8\) reported a DNA extraction procedure from plant tissues using a detergent, CTAB (cetyltrimethylammonium bromide). This method has been widely used not only for plant tissues but for also grains.\(^3\) However it takes 2 days to extract DNA from grain, and a large amount of sample is required. The author reported the use of a simple DNA extraction method\(^15\), but this method needs harmful reagent such as benzyl bromide. The method presented here is based on a freeze-and-thaw cycle that enables to extract purified high molecular weight grain DNA without the use of expensive equipment and/or time-consuming procedures.

In this paper, simple and rapid DNA extraction from milled rice is described, and this procedure was applied to identification of KDML 105 and varieties with similar appearance by microsatellite analysis.

Materials and Methods

1) Samples

Authentic milled Thai rice varieties, Khao Dawk Mali 105, Chainat 1 (CNT1), and RD 7 was obtained from Pathumthani Rice Research Center, Thailand. Two samples, "Khao Dawk Mali" and "Northeastern Khao Dawk Mali", were obtained from markets in Bangkok and Suwannaphum, Thailand, respectively. Malaysian sample, "Beras Wangi Thai", was obtained from market in Kuala Lumpur, Malaysia. For the market samples, 5 grains each were selected randomly and subjected to the identification.

2) Rapid DNA extraction method from single grain of milled rice (freeze-thaw method)

For each sample, a single grain of milled rice was heated at 99 °C with 70μl of ddH\(_2\)O. Heated sample was transferred to a fresh tube, and homogenized in 300μl of 1/10 diluted TE buffer (10mM Tris-HCl 1mM EDTA, pH8.0) with a pellet mixer (Treff 9.7339.9.01). A 400μl aliquot of phenol saturated with
TE buffer was immediately added, the solution was vigorously mixed and incubated in a freezer (-20°C) for 1h. The solution was thawed at room temperature. After centrifugation at 10,000g for 15min, the supernatant was transferred to a fresh tube. An equal volume of phenol:chloroform:3-methyl-1-butanol (50:49:1 v/v/v) was added and the solution was mixed. After centrifugation at 10,000g for 10 min, the supernatant was subjected to DNA precipitation in 1/25 volume of 5M sodium chloride, and 2.5 volumes of ethanol. The DNA pellet was washed twice in 70%(v/v) ethanol, dried at room temperature and resuspended in 30μl of 1/10 TE buffer containing 10mg/ml RNAse.

The presence of genomic DNA was confirmed by electrophoresis on a 0.7%(w/v) agarose gel in TAE buffer (40mM Tris-Acetic acid, 1mM EDTA, pH8.0), stained with ethidium bromide and visualized under UV illumination as well as quantified. Quality was evaluated by absorbance at 260 and 280nm using a SHIMADZU UV-1200 spectrophotometer.

### Table 1. Primer sequences used for cultivar identification of milled rice

<table>
<thead>
<tr>
<th>Locus</th>
<th>Forward</th>
<th>Reverse</th>
</tr>
</thead>
<tbody>
<tr>
<td>OSR6</td>
<td>CCAAGGGAAAAAGATGGCACAA</td>
<td>GTCGACGCTTTATATATGGG</td>
</tr>
<tr>
<td>OSR18</td>
<td>CCCCAAGGAATAATCAGGAGG</td>
<td>TCGGTATGTTTGGTCCCAAG</td>
</tr>
<tr>
<td>OSR22</td>
<td>CTGAGCTCTCTGTTTCCTCATC</td>
<td>CTGTAACTCTGCTGCAC</td>
</tr>
<tr>
<td>OSR28</td>
<td>AGCACTGACACTGCTGAGG</td>
<td>ACTGACATGAGCAGACACA</td>
</tr>
<tr>
<td>OSR29</td>
<td>GCTAGCACTGATAGCTGAC</td>
<td>AGACTGCGCTAGATGACCA</td>
</tr>
<tr>
<td>OSR32</td>
<td>CCTCGAATCCGGCCAAGCTCA</td>
<td>CTTCTGTGACCTGAC</td>
</tr>
<tr>
<td>RM1</td>
<td>GCCGAACACAAATGCACAA</td>
<td>GCTGTTGACCCCTGAC</td>
</tr>
<tr>
<td>RM4</td>
<td>TTAGCACGATCGACACTGAC</td>
<td>AAGGTGTAACCCACTGAC</td>
</tr>
<tr>
<td>RM10</td>
<td>TTTGTCAAGGAGGAGCATCG</td>
<td>CAGATGGAAGATGAGTCC</td>
</tr>
<tr>
<td>RM16</td>
<td>CGCTAGGGCAATGCTGAAA</td>
<td>AACAGCAGTACGCCCG</td>
</tr>
<tr>
<td>RM17</td>
<td>TGCCCTGTATTCTTTCTCTC</td>
<td>GGTGATCCTTTCCATTTCA</td>
</tr>
<tr>
<td>RM19</td>
<td>CAAAACACAGGAGCATAGAC</td>
<td>CTCAAGATGGACCCGAAG</td>
</tr>
<tr>
<td>RM122</td>
<td>GAGGTGATCAATAGCATCAGTGC</td>
<td>GAAGGAATGACAGCCAG</td>
</tr>
</tbody>
</table>

3) Microsatellite analysis (PCR)

Selected primers, flanking the tandem repeats, shown in Table 1, were used for amplification of genomic DNA fragments. PCR amplifications were performed in a total volume of 20μl containing 3μl of extract, 0.1mmol of each primer, 200mM each of dNTP, 50mM KCl, 10mM Tris-HCl (pH8.3), 0.01% gelatin, 3mM MgCl₂ and 0.5unit rTaq DNA polymerase (TOYOBO). PCR was initiated by a denaturation step at 94°C for 2min, followed by 35 cycles of 55°C for 1min, 72°C for 1min, and a final extension at 72°C for 5min. PCR was performed on a Corbett Research PC-960G. Amplification products were resolved by electrophoresis on a 3% agarose gel in TAE buffer, stained with ethidium bromide and visualized under UV illumination.

The reactions were repeated at least 3 times to confirm the reproducibility of the results.

### Results and Discussion

1) Rapid DNA extraction from single grain of milled rice (freeze-thaw method)

The purity of the DNA, determined from the A₂₆₀/A₂₈₀ ratio averaged > 1.78 for all the samples. There was no evidence of degrading of DNA during preparation (Fig. 1). The yield of DNA ranged from 200 to 300 ng/single grain for all the samples. Since a sample of milled rice was once heated to break the cell wall and to denature endogenous nucleases in the method (Fig. 1). Polysaccharides, inhibiting the enzyme reaction, were removed efficiently during the freezing and thawing cycle.

2) Identification of variety by using microsatellite analysis

The primer set RM17 was particularly useful for discriminating the 3 varieties (Fig. 2). The market samples were analyzed by RM17.

Fig. 3 shows the results of microsatellite analysis of market samples on agarose gel. "Beras Wangi Thai", sampled in a Kuala Lumpur market, and "Khao Dawk Mali", in a Bangkok market, did not correspond to Khao Dawk Mali 105 variety. Khao Dawk Mali 105 was identified only from "Northeastern
Khao Dawk Mali”, sampled in a Suwannaphum market.

A simple and rapid method of cultivar identification of milled rice is illustrated in Fig. 4. By applying the improved method, we were able to obtain DNA for microsatellite analysis within 2 hr and identification required 6 hr in total.

The efficiency, speed and the lack of expensive instruments or toxic chemicals in the present method are an attractive alternative to the existing methods of rice grain genomic DNA. These results show that DNA extracted in this simple, low-cost, rapid and safe protocol is of high quality and can be used in PCR-based techniques for cultivar identification even in laboratories with a moderate level of technology.

Acknowledgements

The author thanks Ms. Ngamchuen Kongseree, Pathumthani Rice Research Center, Rice Research Institute, Thailand, and Dr. Nobuyuki Kabaki, JIRCAS, Khon Kaen, Thailand for the supply of authentic and market samples.

This study was carried out within the framework of the North East Thailand Project sponsored by
Fig. 4. Scheme of rapid DNA extraction and cultivar identification of milled rice.
REFERENCES


精米からの簡易迅速 DNA 抽出法の開発と、
そのカオドマリ 105 への応用

吉橋 忠

国際農林水産業研究センター生産利用部
（〒 305-8686 茨城県つくば市大わし１－２）

摘 要

香り米品種カオドマリ 105 はタイにおける主要な品種である。しかしながら、生産量の拡大はカオドマリの種々の特性により困難である。さらに、客観的な評価法がいないため不正な取引が野放しになっている。そのため、客観的な品種判別技術の開発が必要とされている。PCR 法をベースとした分子マーカー法は客観的な品種判別法として有用であるが、その適用には、高分子量の DNA を精米１粒１粒から抽出する必要がある。筆者は凍結・融解を利用し、迅速かつ高価な機器が必要でない高分子量の DNA の抽出法を開発した。抽出した DNA のマイクロサテライト分析を行うことにより、品種判別を行った。バンコク・スワンナプーム・クアラルンプールそれぞれの市場のサンプルを分析することにより、バンコク・クアラルンプール市場品より、カオドマリ 105 ではない品種を検出した。本抽出法を品種判別に適用することにより、DNA 抽出まで２時間、品種判別まで 6 時間以内に判別することが可能となった。

キーワード：品種判別、PCR、迅速抽出、ゲノム DNA