REVIEW

Cross-tolerance to Thermal Stresses and its Application to the Development of Cold Tolerant Rice

Hiroshi YASUDA*

Crop Breeding Research Division, Hokkaido Agricultural Research Center, NARO (Sapporo, Hokkaido 062-8555, Japan)

Abstract

Having originated in tropical regions, rice is more sensitive to cold stress than other grain crops, such as rye, wheat, and barley, which originated in temperate regions. Despite the limited knowledge about the exact mechanisms of cold tolerance, breeders have generated new rice cultivars that can grow in Hokkaido, Japan's northern limit of rice cultivation, for more than a hundred years. However, greater cold tolerance in rice would enable more stable production and elevate productivity. Because cold tolerance is a complex quantitative trait, it is difficult to pyramid the genes that could improve plant tolerance to this abiotic stress. Therefore, it is important to develop effective strategies for improving cold tolerance in rice. In this review, the use of a 'cross-tolerance' strategy is proposed, and the current status of such strategy introduced.

Discipline: Biotechnology, Crop production

Additional key words: abiotic stress, reactive oxygen species, heat shock factor

Introduction

The expansion of areas for rice production is important for world food security in the face of significant population growth, particularly in Asia and Africa. Hokkaido, in northern Japan, represents the northern limit for the economical production of rice. Rice breeders in Hokkaido must contend with cool summers and relatively short durations of sunshine during rice heading. However, rice growers in the area still face such problems as ensuring the cropping season and avoiding chilling damage.

There are two major critical periods for the cultivation of rice in cold regions. These include the stage from germination through seedling growth and the booting stage. Low temperatures adversely affect germination and early growth, as rice seedlings are sensitive to cold. Therefore, farmers grow rice seedlings in greenhouse nurseries to ensure germination and adequate early development of the seedlings. Later, cold stress at the booting stage inhibits pollen development, which leads to reduced fertility and yield. Cold tolerance at the booting stage is a more important issue as it directly affects farmers' incomes. Over the past few decades, conventional breeding has generated rice cultivars with cold tolerance at the booting stage, thereby

gradually improving cold tolerance. The adverse effects of cold stress have thus been avoided at early stages by a special cultivation technique and at the booting stage by the effective use of genetic resources.

Global warming can certainly increase the incidence of extreme weather patterns and events, such as hot and cool summers or localized torrential rains. Even now, plant cold tolerance is one of the most important agricultural problems to be addressed in the Hokkaido region. Due to their immobility, plants have evolved several molecular mechanisms to withstand abiotic stresses in the environment. Therefore, the mechanisms of tolerance to abiotic stresses due to the changeable environment should be investigated in detail, with the resulting information being used to create more stable and higher-yielding crops.

We have been conducting experiments designed to control the cold tolerance of rice at two critical periods: from germination through the seedling stage, and at the booting stage. The results of these experiments suggest that cold tolerance at both stages might be enhanced by the over-expression of reactive oxygen species (ROS)-scavenging enzymes and activation of the heat shock response under cold conditions. In this review, we discuss the relationship between cold tolerance, ROS, and the heat shock response,

and introduce strategies to mitigate cold stress by exploiting these pathways.

Response to abiotic stress

1. Response to cold

When plants that originated in tropical regions are exposed to low temperatures below 10-12°C, they can experience chilling damage due to physiological dysfunction (Lyons 1973). Low temperatures in Australia reduce the average annual rice yield by 5-10%, and unpredictable cold snaps can result in yield losses of 20-40% (Oliver et al. 2005). In Japan, similar yield losses are caused by unpredictable cold snaps that occur roughly every three to four years.

As plants are sessile, they have evolved morphological and physiological responses to abiotic stress involving genes in stress-responsive pathways in order to survive or increase their range of distribution. Although unique sensors for low temperature have yet to be identified, plant cells may sense cold stress via changes in the fluidity of cellular membranes (Yadav 2010, Miura & Furumoto 2013). These changes in the consistency of cell membranes can result in increased cytosolic calcium levels (Knight et al. 1991). These signals can then induce such cold-stress responsive genes as OsICE1/2 and activate the DREB-CRT/DRE pathway (Zhu et al. 2007, Nakamura et al. 2011).

Many genes responsive to cold temperatures including chilling (0-15°C) and freezing (below 0°C) conditions have been identified in the last two decades. The functions of cold-responsive genes and QTL in the cold tolerance of rice (Table 1) have been reviewed and summarized by da Cruz et al. (2013) and Zhang et al. (2014). The expression and function of genes involved in stress response are controlled by stress-inducible transcription factors (TFs) such as DREB. Because DREB is one of the most important TFs under cold-stress conditions, the DREB pathway is a powerful tool for conferring cold tolerance to crops (Ito et al. 2006). Other TFs such as NAC and MYB can also influence cold tolerance, as can CDPK and MAPK, which are involved in signal transduction. These results suggest that the enhancement of cold tolerance requires harmonization of the expression and function of several genes, including genes that affect TFs and the genes controlled by TFs.

2. Thermal stress and ROS generation

Abiotic stresses such as heat, cold, drought, salinity, and ozone generally accelerate the generation of ROS such as hydrogen peroxide, the hydroxyl radical, and superoxide (Mittler 2002, Apel & Hirt 2004, Suzuki & Mittler 2006, Miller et al. 2008). Although cold stress signaling in plant cells can be controlled by ROS accumulation as a second messenger, excessive levels of ROS lead to cell death

(Sharma et al. 2012). Therefore, when plant cells are exposed to any abiotic stress condition, the expression of reactive oxygen-scavenging enzymes is usually elevated. Thus, different stress conditions can induce the same ROS response, which suggests that the homeostasis of redox balance in stressed cells is very important.

When rice seedlings were pre-treated under heat shock conditions (42°C), the seedlings became more tolerant to cold. Although the mechanism is not completely understood, overexpression of the ascorbate peroxidase (APX) gene, which is induced by high temperature, can confer cold tolerance (Sato et al. 2001). APX is a typical ROS-scavenging enzyme that catalyzes the conversion of H₂O₂ to its detoxified form H₂O. Further, overexpression of DHAR (dehydroascorbate reductase), which is indirectly related to the detoxification of ROS, can also confer cold tolerance to rice (Yasuda et al. unpublished data). These results suggest that enhancing ROS-scavenging ability could be an effective strategy for enhancing plant tolerance to cold stress.

Heat stress also induces the production of excess amounts of ROS in plant cells; therefore, the activities of many ROS-scavenging enzymes in several plant species are elevated under heat stress conditions (Kumar et al. 2012, Lin et al. 2010). Miller and Mittler (2006) found that ROS induces the expression of genes encoding heat shock factors (HSFs) and Davletova et al. (2005) described the central role of HSFs in sensing H₂O₂ stress. These reports indicated that HSFs play a pivotal role in mitigating excess ROS.

HSFs are known to be transcription factors that induce the expression of HSPs, and play a central role in the heat shock response. HSFs are encoded by plant, animal, and bacterium genomes, but plant HSFs are more diverse than those in other kingdoms. For instance, HSFs in plants are encoded by multi-gene families and the *Arabidopsis*, rice, and tobacco genomes each encode at least 21, 25, and 17 *Hsf* genes, respectively (Baniwal et al. 2004, Guo et al. 2008, Mishra et al. 2002, Mittal et al. 2009, von Koskull-Döring et al. 2007). Conversely, mammals, *Chlamidomonas*, yeast, and *Drosophila* have only three genes, two genes, one gene, and one Hsf gene, respectively (Morimoto 1998, Schulz-Raffelt et al. 2007). Taken together, this suggests that plants have a complex and highly regulated system for heat shock response.

In general, HSFs are classified into three classes (A, B, and C) based on domain structure. The activation function in transcriptional activators only has the A-type of HSF, because the activation domain only exists in class A HSFs (Kotak et al. 2004, Nover et al. 2001). Members of the *Hsf* gene family have various functions; for example, HSFA1 in tomato and *Arabidopsis* acts as a master regulator of the heat stress response (Mishra et al. 2002, Liu et al. 2011), HSFA2 in rice and *Arabidopsis* confers tolerance to

JARQ 51 (2) 2017

heat shock, high light, oxidative, salt, and anoxia stresses (Charng et al. 2007, Ogawa et al. 2007, Yokotani et al. 2008, Banti et al. 2010), HSFA3 in *Arabidopsis* influences drought stress tolerance (Sakuma et al. 2006, Yoshida et al. 2008), and HSFA4 in wheat and *Arabidopsis* confers tolerance to cadmium (Shim et al. 2009) and salt (Perez-Salamo et al. 2014). Thus, HSFs have been implicated in tolerance to a wide range of abiotic stresses. Expression of HSFA3 in rice is also induced by low temperature (Nakamura et al. 2011), but there is no evidence that HSFs confer cold stress tolerance.

3. Cross-tolerance to multiple stresses

An interesting phenotype known as cross-tolerance, in which exposure to one type of stress condition confers tolerance to other types of stress, is a phenomenon commonly observed in plants (Bartoli et al. 2013). Table 1 lists the useful genes that can confer stress tolerance to multiple stresses in rice, when the expression and function of the genes are activated. Most of these genes encode TFs or enzymes involved in the synthesis of compatible solutes. Because TFs can simultaneously induce many downstream genes that act to acquire abiotic stress tolerance, using TF has an advantage compared to using a single functional

Table 1. List of genes which coincidently enhanced multiple stress tolerance in rice

Gene	Origin of gene	Function	Tolerance	Reference
Alsap	Aeluropus littoralis	Transcription control	cold, drought, salt	Saad et al. 2012
bZIP23	rice	Transcription control	drought, salt	Xiang et al. 2008
CBF	maize	Transcription control	cold, drought, salt	Xu et al. 2011
COIN	rice	Transcription control	cold, drought, salt	Liu et al. 2007
DREB1s	rice	Transcription control	cold, drought, salt	Ito et al. 2006
DREB1F	rice	Transcription control	cold, drought, salt	Wang et al. 2008
MYB2	rice	Transcription control	cold, drought, salt	Yang et al. 2012
MYB48-1	rice		drought, salt	Xiong et al. 2014
NAC5	rice	Transcription control	cold, drought, salt	Song et al. 2011
SNAC2	rice	Transcription control	cold, drought, salt	Hu et al. 2008
WRKY11	rice	Transcription control	drought, heat	Wu et al. 2009
ZFPs	rice	Transcription control	cold, drought, salt	Huang et al. 2009, 2012, Xu et al. 2008
ADC	Datura stramonium oat	Synthesis of polyamine	drought, salt	Capell et al. 2004 Roy & Wu 2001
codA	Arthrobacter globiformis	Synthesis of glycinebetaine	cold, salt	Sakamoto et al. 1998
P5CS	moth bean	Synthesis of proline	drought, salt	Zhu et al. 1998
TPSP TPS1	Escherichia coli rice	Synthesis of trehalose	cold, drought, salt	Garg et al. 2002 Li et al. 2011
HVA1	barley	LEA protein	drought, salt	Xu et al. 1996
LEA4	rice	LEA protein	drought, salt, heavy metal	Hu et al. 2016
HSP17.0	rice	Molecular chaperon	drought, salt	Zou et al. 2011
HSP23.7	rice			
HSP18.6	rice	Molecular chaperon	cold, drought, salt, heat	Wang et al. 2015
sHSP17.7	rice	Molecular chaperon	heat, UV-B, drought	Murakami et al. 2004, Sato & Yokoya 2008
CDPK7	rice	Ca ²⁺ -dependent protein kinase	cold, drought, salt	Saijo et al. 2000
MAPK5	rice	Mitogen-activated protein kinase	cold, drought, salt	Xiong & Yang 2003
PYL5	rice	Hormone receptor	drought, salt	Kim et al. 2014
TaSTRG	wheat	Salt-induced unknown	drought, salt	Zhou et al. 2009

gene. However, the transformants with several introduced TF genes might represent such off-target effects as dwarfism and retarded growth, when expression of the TF was enhanced under normal conditions (Ito et al. 2006, Wu et al. 2009, Xu et al. 2011).

The expression of most TFs listed in Table 1 is enhanced by stress conditions (Sakuma et al. 2002, Agarwal et al. 2007, Saad et al. 2010, Takasaki et al. 2010). TFs sometimes respond to the presence of ROS such as H₂O₂ via protein kinases, through Ca²⁺ as a second messenger, a mechanism common to plant responses to several abiotic stresses (Wang et al. 2003). These results suggest that intracellular ROS could induce molecular cascades that lead to abiotic stress tolerance. This phenomenon could thus form the basis of a strategy for generating abiotic stress-tolerant crops.

Utilization of HSFs for enhancing cold stress tolerance

Taken together, the results above raise the question of why the heat shock response could confer cold tolerance to rice, for several reasons. First, some heat-shock inducible genes in rice and *Brassica napus* such as low-molecular-mass HSP (Wang et al. 2015), high-molecular-mass HSPs

(Krishna et al. 1995, Pareek et al. 1995), and ascorbate peroxidase (Sato et al. 2001) are upregulated under cold stress. Secondly, heat shock factors function as H_2O_2 sensors (Miller and Mittler 2006) and heat shock responsive genes have been implicated in ROS equilibrium (Davletova et al. 2005, Kumar et al. 2012).

To activate the heat shock response under cool conditions, we used a cold-responsive wheat promoter (WCR promoter) to control the expression of HsfA2 in rice (Yasuda et al. 2013). However, the heat shock response was not clearly activated in the transformants, even though they overexpressed the transgenes under cool conditions. Therefore, an attempt was made for the co-expression in rice of HsfA2, together with the dominant negative form of HSP90 that controls the HsfA2 function. HSP90 is a negative regulator of HSF, and inhibition of HSP90 is required for the induction of heat-inducible genes (Yamada & Nishimura 2007). When the rice transformants co-expressed both the gene encoding HsfA2 and the gene encoding the dominant negative form of HSP90, the heat shock response was activated in rice under cool conditions (Yasuda et al. 2013). These results suggest that the heat shock response is strongly regulated in plant cells (Fig. 1). Although this experiment is ongoing, so far these rice transformants have improved cold stress tolerance during the seedling stage

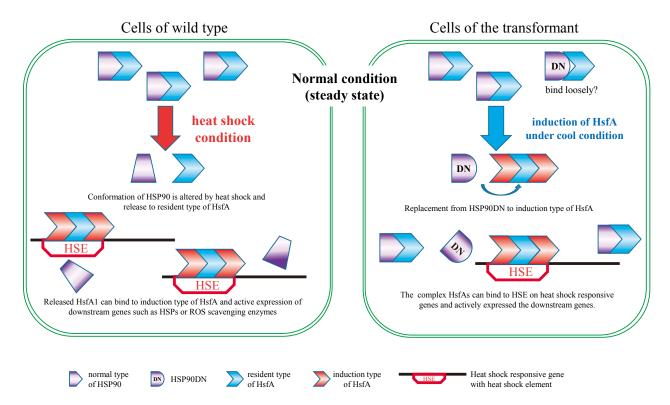


Fig. 1. A model for activation of rice heat shock response (HSR).

Left: induction of HSR under heat shock condition in wild type cells; Right: induction of HSR under cool condition in the transformants described by Yasuda et al. (2013).

JARQ 51 (2) 2017

(Yasuda et al. unpublished data).

Future perspectives

Due to the rapid advance of climate change caused by global warming, crops tolerant to abiotic stresses should be generated as quickly as possible. Climate change not only involves warming but is also predicted to induce drought or cold locally, which would have significant effects on crop yields. Table 1 shows that one gene conferred multiple forms of stress tolerance to rice; whereas most of the genes confer tolerance to cold, drought, and salt. However, the mechanisms of this tolerance have yet to be completely elucidated, and most of the genes listed are involved in the synthesis of osmolytes or are chaperones, except for the transcription factor. These results suggest that these three stresses induce similar cellular damage in response to osmotic and/or oxidative stress (Wang et al. 2003), and activate similar signal pathways for maintaining homeostasis of the cellular environment under stress (Zhu et al. 2002). If it were possible to completely control the activity of these genes, we would be able to grow complete crops highly tolerant to multiple severe stress conditions, including cold. It is necessary to develop new crops with greater tolerance to multiple abiotic stress and higher or at least stable yield under stress, in order to feed the world's growing population. Thus, cross-tolerance should be a key word for the next generation of crop breeding.

References

- Agarwal, P. et al. (2007) Genome-wide identification of C₂H₂ zinc-finger gene family in rice and their phylogeny and expression analysis. *Plant Mol. Biol.*, **65**, 467-485.
- Apel, K. and Hirt, H. (2004) Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *Annu. Rev. Plant Biol.*, **55**, 373-399.
- Baniwal, S. K. et al. (2004) Heat stress response in plants: a complex game with chaperones and more than twenty heat stress transcription factors. *J. Biosci.*, **29**, 471-487.
- Banti, V. et al. (2010) The heat-inducible transcription factor *HsfA2* enhances anoxia tolerance in Arabidopsis. *Plant Physiol.*, **152**, 1471-1483.
- Bartoli, C. G. et al. (2013) Interactions between hormone and redox signaling pathways in the control of growth and cross tolerance to stress. *Environ. Exp. Bot.*, **94**, 73-88.
- Capell, T. et al. (2004) Modulation of the polyamine biosynthetic pathway in transgenic rice confers tolerance to drought stress. *Proc. Natl. Acad. Sci. USA*, **101**, 9909-9914.
- Charng, Y. Y. et al. (2007) A heat-inducible transcription factor, HsfA2, is required for extension of acquired thermotolerance in Arabidopsis. *Plant Physiol.*, **143**, 251-262.
- da Cruz, R. P. et al. (2013) Avoiding damage and achieving cold

- tolerance in rice plants. Food Energy Secur., 2, 96-119.
- Davletova, S. et al. (2005) Cytosolic ascorbate peroxidase 1 is a central component of the reactive oxygen gene network of Arabidopsis. *Plant Cell*, **17**, 268-281.
- Garg, A. K. et al. (2002) Trehalose accumulation in rice plants confers high tolerance levels to different abiotic stresses. *Proc. Natl. Acad. Sci. USA*, 99, 15898-15903.
- Guo, J. et al. (2008) Genome-wide analysis of heat shock transcription factor families in rice and *Arabidopsis. J. Genet. Genomics*, **35**, 105-118.
- Hu, H. et al. (2008) Characterization of transcription factor gene *SNAC2* conferring cold and salt tolerance in rice. *Plant Mol. Biol.*, **67**, 169-181.
- Hu, T. et al. (2016) Overexpression of OsLEA4 enhances drought, high salt and heavy metal stress tolerance in transgenic rice (Oryza sativa L.). Environ. Exp. Bot., 123, 68-77.
- Huang, J. et al. (2009) Increased tolerance of rice to cold, drought and oxidative stresses mediated by the overexpression of a gene that encodes the zinc finger protein ZFP245. *Biochem. Biophys. Res. Commun.*, 389, 556-561.
- Huang, J. et al. (2012) A TFIIIA-type zinc finger protein confers multiple abiotic stress tolerances in transgenic rice (*Oryza sativa* L.). *Plant Mol. Biol.*, **80**, 337-350.
- Ito, Y. et al. (2006) Functional analysis of rice DREB1/CBF-type transcription factors involved in cold-responsive gene expression in transgenic rice. *Plant Cell Physiol.*, **47**, 141-153.
- Kim, H. et al. (2014) Overexpression of *PYL5* in rice enhances drought tolerance, inhibits growth, and modulates gene expression. *J. Exp. Bot.*, **65**, 453-464.
- Knight, M. R. et al. (1991) Transgenic plant aequorin reports the effects of touch and cold-shock and elicitors on cytoplasmic calcium. *Nature*, 252, 524-526.
- Kotak, S. et al. (2004) Characterization of C-terminal domains of *Arabidopsis* heat stress transcription factors (Hsfs) and identification of a new signature combination of plant class A Hsfs with AHA and NES motifs essential for activator function and intracellular localization. *Plant J.*, 29, 98-112.
- Krishna, P. et al. (1995) Cold-induced accumulation of hsp90 transcripts in *Brassica napus*. *Plant Physiol.*, **107**, 915-923.
- Kumar, S. et al. (2012) Comparative response of maize and rice genotypes to heat stress: status of oxidative stress and antioxidants. *Acta. Physiol. Plant.*, **34**, 75-86.
- Li, H. W. et al. (2011) Overexpression of the trehalose-6-phosphate synthase gene *OsTPS1* enhances abiotic stress tolerance in rice. *Planta*, **234**, 1007-1018.
- Lin, K. H. et al. (2010) Cloning, expression and physiological analysis of broccoli catalase gene and Chinese cabbage ascorbate peroxidase gene under heat stress. *Plant Cell Rep.*, 29, 575-593.
- Liu, H. C. et al. (2011) The role of class A1 heat shock factors (HSFA1s) in response to heat and other stresses in *Arabidopsis*. *Plant Cell Environ.*, **34**, 738-751.
- Liu, K. et al. (2007) Overexpression of OsCOIN, a putative cold

- inducible zinc finger protein, increased tolerance to chilling, salt and drought, and enhanced proline level in rice. *Planta*, **226**, 1007-1016.
- Lyons, J. M. (1973) Chilling injury in plants. *Ann. Rev. Plant Physiol.*, **24**, 445-466.
- Miller, G. and Mittler, R. (2006) Could heat shock transcription factors function as hydrogen peroxide sensors in plants? *Ann. Bot.*, **98**, 279-288.
- Miller, G. et al. (2008) Reactive oxygen signaling and abiotic stress. *Physiol. Plant.*, **133**, 481-489.
- Mishra, S. K. et al. (2002) In the complex family of heat stress transcription factors, HsfA1 has a unique role as master regulator of thermotolerance in tomato. *Genes. Dev.*, 16, 1555-1567.
- Mittal, D. et al. (2009) Heat shock factor gene family in rice: Genomic organization and transcript expression profiling in response to high temperature, low temperature and oxidative stresses. *Plant Physiol. Biochem.*, 47, 785-795.
- Mittler, R. (2002) Oxidative stress, antioxidants and stress tolerance. *Trends Plant Sci.*, 7, 405-410.
- Miura, K. and Furumoto, T. (2013) Cold signaling and cold response in plants. *Int. J. Mol. Sci.*, **14**, 5312-5337.
- Morimoto, R. (1998) Regulation of the heat shock transcriptional response: cross talk between a family of heat shock factors, molecular chaperones, and negative regulators. *Genes Dev.*, **12**, 3788-3796.
- Murakami, T. et al. (2004) Over-expression of a small heat shock protein, sHSP17.7, confers both heat tolerance and UV-B resistance to rice plants. *Mol. Breed.*, 13, 165-175.
- Nakamura, J. et al. (2011) Rice homologs of inducer of *CBF* expression (OsICE) are involved in cold accumulation. *Plant Biotechnol.*, **28**, 303-309.
- Nover, L. et al. (2001) *Arabidopsis* and the heat stress transcription factor world: how many heat stress transcription factors do we need? *Cell Stress Chaperones*, **6**, 177-189.
- Ogawa, D. et al. (2007) High-level overexpression of the *Arabidopsis HsfA2* gene confers not only increased thermotolerance but also salt/osmotic stress tolerance and enhanced callus growth. *J. Exp.Bot.*, **58**, 3373-3383.
- Oliver, S. N. et al. (2005) Cold-induced repression of the rice anther-specific cell wall invertase gene *OSINV4* is correlated with sucrose accumulation and pollen sterility. *Plant Cell Environ.*, **28**, 1534-1551.
- Pareek, A. et al. (1995) Immunological evidence for accumulation of two high-molecular-weight (104 kDa and 90 kDa) HSPs in response to different stresses in rice and in response to high temperature stress in diverse plant genera. *Plant Mol. Biol.*, **29**, 293-301.
- Perez-Salamo, I. et al. (2014) The teat shock factor A4A confers salt tolerance and is regulated by oxidative stress and the mitogen-activated protein kinases MPK3 and MPK6. *Plant Physiol.*, **165**, 319-334.
- Roy, M. and Wu, R. (2001) Arginine decarboxylase transgene

- expression and analysis of environmental stress tolerance in transgenic rice. *Plant Sci.*, **160**, 869-875.
- Saad, R. B. et al. (2010) Improved drought and salt stress tolerance in transgenic tobacco overexpressing a novel A20/AN1 zinc-finger "AISAP" gene isolated from the halophyte grass Aeluropus littoralis. Plant Mol. Biol., 72, 171-190.
- Saad, R. B. et al. (2012) Expression of the *Aeluropus littoralis AISAP* gene in rice confers broad tolerance to abiotic stresses through maintenance of photosynthesis. *Plant Cell Environ.*, **35**, 626-643.
- Sabehat, A. et al. (1996) The correlation between heat-shock protein accumulation and persistence and chilling tolerance in tomato fruit. *Plant Physiol.*, 110, 531-537.
- Saijo, Y. et al. (2000) Over-expression of a single Ca²⁺-dependent protein kinase confers both cold and salt/drought tolerance on rice plants. *Plant J.*, **23**, 319-327.
- Sakamoto, A. et al. (1998) Metabolic engineering of rice leading to biosynthesis of glycinebetaine and tolerance to salt and cold. *Plant Mol. Boil.*, **38**, 1011-1019.
- Sakuma, Y. et al. (2002) DNA-binding specificity of the ERF/AP2 domain of *Arabidopsis* DREBs, transcription factors involved in dehydration- and Cold-inducible gene expression. *Biochem. Biophys. Res. Comm.*, 290, 998-1009.
- Sakuma, Y. et al. (2006) Dual function of an *Arabidopsis* transcription factor DREB2A in water-stress-responsive and heat-stress-responsive gene expression. *Proc. Natl. Acad. Sci. USA*, 103, 18822-18827.
- Sato, Y. and Yokoya, S. (2008) Enhanced tolerance to drought stress in transgenic rice plants overexpressing a small heatshock protein, sHSP17.7. *Plant Cell Rep.*, 27, 329-334.
- Sato, Y. et al. (2001) Heat shock-mediated APX gene expression and protection against chilling injury in rice seedling. *J. Exp. Bot.*, **52**, 145-151.
- Schulz-Raffelt, M. et al. (2007) Heat shock factor 1 is a key regulator of the stress response in *Chlamydomonas*. *Plant J.*, **52**, 286-295.
- Sharma, P. et al. (2012) Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. *J. Bot.*, **2012**, 217037.
- Shim, D. et al. (2009) Orthologs of the class A4 heat shock transcription factor HsfA4a confer cadmium tolerance in wheat and rice. *Plant Cell*, **21**, 4031-4043.
- Song, S. Y. et al. (2011) Physiological mechanisms underlying OsNAC5-dependent tolerance of rice plants to abiotic stress. *Planta*, **234**, 331-345.
- Suzuki, N. and Mittler, R. (2006) Reactive oxygen species and temperature stresses: A delicate balance between signaling and destruction. *Physiol. Plant.*, 126, 45-51.
- Takahashi, H. et al. (2010) The abiotic stress-responsive NAC-type transcription factor OsNAC5 regulates stress-inducible genes and stress tolerance in rice. *Mol. Genet. Genomics*, **284**, 173-183.
- van Berkel, J. et al. (1994) Transcripts accumulating during

JARQ 51 (2) 2017

- cold storage of potato (*Solanum tuberosum* L.) tubers are sequence related to stress-responsive genes. *Plant Physiol.*, **104**, 445-452.
- von Koskull-Döring, P. et al. (2007) The diversity of plant heat stress transcription factors. *Trends Plant Sci.*, **12**, 452-457.
- Wang, A. et al. (2015) Overexpression of a small heat-shockprotein gene enhances tolerance to abiotic stresses in rice. *Plant Breed.*, 134, 384-393.
- Wang, H. et al. (1995) Promoters from kin1 and cor6.6, two homologous Arabidopsis thaliana genes: transcriptional regulation and gene expression induced by low temperature, ABA, osmoticum and dehydration. Plant Mol. Biol., 28, 605-617.
- Wang, Q. et al. (2008) Overexpression of a rice *OsDREB1F* gene increases salt, drought, and low temperature tolerance in both *Arabidopsis* and rice. *Plant Mol. Biol.*, **67**, 589-602.
- Wang, W. et al. (2003) Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance. *Planta*, **218**, 1-14.
- Wu, X. et al. (2009) Enhanced heat and drought tolerance in transgenic rice seedlings overexpressing *OsWRKY11* under the control of *HSP101* promoter. *Plant Cell Rep.*, 28, 21-30.
- Xiang, Y. et al. (2008) Characterization of OsbZIP23 as a key player of the basic leucine zipper transcription factor family for conferring abscisic acid sensitivity and salinity and drought tolerance in rice. *Plant Physiol.*, 148, 1938-1952.
- Xiong, H. et al. (2014) overexpression of *OsMYB48-1*, a novel MYB-related transcription factor, enhances drought and salinity tolerance in rice. *PLoS One*, 9, e92913.
- Xiong, L. and Yang Y. (2003) Disease resistance and abiotic stress tolerance in rice are inversely modulated by an abscisic acid-inducible mitogen-activated protein kinase. *Plant Cell*, 15, 745-759.
- Xu, D. et al. (1996) Expression of a late embryogenesis abundant protein gene, *HVA1*, from barley confers tolerance to water deficit and salt stress in transgenic rice. *Plant Physiol.*, **110**, 249-257.
- Xu, D. Q. et al. (2008) Overexpression of a TFIIIA-type zinc fin-

- ger protein gene *ZFP252* enhances drought and salt tolerance in rice (*Oryza sativa* L.). *FEBS Lett.*, **582**, 1037-1043.
- Xu, M. et al. (2011) ZmCBF3 overexpression improves tolerance to abiotic stress in transgenic rice (*Oryza sativa*) without yield penalty. *Plant Cell Rep.*, 30, 1949-1957.
- Yadav, S. K. (2010) Cold stress tolerance mechanisms in plants. A review. Agron. Sustain. Dev., 30, 515-527.
- Yang, A. et al. (2012) A R2R3-type MYB gene, OsMYB2, is involved in salt, cold, and dehydration tolerance in rice. *J. Exp. Bot.*, **63**, 2541-2556.
- Yasuda, H. et al. (2013) Generation of transgenic rice expressing heat shock protein genes under cool conditions. *Plant Biotechnol.*, 30, 489-496.
- Yokotani, N. et al. (2008) Expression of rice heat stress transcription factor OsHsfA2e enhances tolerance to environmental stresses in transgenic *Arabidopsis*. *Planta*, 227, 957-967.
- Yoshida, T. et al. (2008) Functional analysis of an *Arabidopsis* heat-shock transcription factor *HsfA3* in the transcriptional cascade downstream of the DREB2A stress-regulatory system. *Biochem. Biophys. Res. Commun.*, **368**, 515-521.
- Zhang, Q. et al. (2014) Rice and cold stress: methods for its evaluation and summary of cold tolerance-related quantitative trait loci. *Rice*, 7, 24.
- Zhou, W. et al. (2009) Overexpression of *TaSTRG* gene improves salt and drought tolerance in rice. *J. Plant Physiol.*, **166**, 1660-1671.
- Zhu, B. et al. (1998) Overexpression of a Δ¹-pyrroline-5-carboxylate synthetase gene and analysis of tolerance to water- and salt-stress in transgenic rice. *Plant Sci.*, **139**, 41-48.
- Zhu, J. et al. (2007) Interplay between cold-responsive gene regulation, metabolism and RNA processing during plant cold acclimation. *Curr. Opin. Plant Biol.*, 10, 290-295.
- Zhu, J. K. (2002) Salt and drought stress signal transduction in plants. *Annu. Rev. Plant Biol.*, **53**, 247-273.
- Zou, J. et al. (2012) Overexpression of *OsHsp17.0* and *OsHsp23.7* enhances drought and salt tolerance in rice. *J. Plant Physiol.*, **169**, 628-635.