

Switch in Searching Behavior and Stimulus Causing Host Examination by *Trichogramma chilonis* Ishii, an Egg Parasitoid of *Plutella xylostella* L.

Kazuki MIURA and Masahiro KOBAYASHI

Department of Agro-Environmental Management, Chugoku National Agricultural Experiment Station (Fukuyama, Hiroshima, 721 Japan)

Abstract

The locomotory response of walking *Trichogramma chilonis* female to the contact of scales and secretions of host moth, *Plutella xylostella*, and elicitation of oviposition behavior were observed. In response to a contaminated Sealon film with host moth scales and secretions, a *T. chilonis* female exhibited a response involving walking at a reduced speed. *T. chilonis* female remained during a much longer period of time on a contaminated Sealon film than on a clean one. Also moth scales and/or secretions elicited the oviposition behavior of *T. chilonis* females.

Discipline: Insect pest

Additional key words: biological control, moth scales and secretions

Introduction

The diamondback moth (DBM), *Plutella xylostella* (L.), is a noted defoliator of numerous cruciferous plants in many areas of the world. Since the resistance to many of the commonly used insecticides makes it one of the most difficult pests to manage^{1,16}, it was necessary to develop alternative methods of control that did not depend on toxic agents, including biological control. *Trichogramma chilonis* Ishii is an important solitary egg parasitoid of DBM^{9,25}. Since much of the work in biological control may be characterized as "trial-error" methods¹⁴, it is important to predict how released parasitoids might behave. Although a large number of studies has been carried out on various aspects of the biology of *T. chilonis*¹⁷⁻²⁰, relatively little is known about how released *T. chilonis* females search for DBM eggs and which cues elicit the oviposition behavior on DBM eggs. The present study was carried out to analyze the searching behavior and elicitation of the oviposition behavior of *T. chilonis* females on DBM eggs in the laboratory.

Materials and methods

T. chilonis used was a part of the stock culture

stored at the Chugoku National Agricultural Experiment Station, which was derived from the original colony introduced from Taiwan⁸ and reared with *Ephestia kuehniella* Zellar eggs under 24°C and 16L-8D conditions. Larvae and pupae of DBM were collected from cabbage fields in Fukuyama, Hiroshima Prefecture, in 1991. The stock culture of the DBM was continuously maintained in a cabinet controlled at 24°C and cabbage leaves were supplied as a food for the larvae. The moths emerging were released into a rearing cage (34 × 34 × 26 cm) containing a piece of Sealon film[®] (Fuji Photo Film Co., LTD) as a substratum for oviposition and honey solution (20%) as food.

Experiments were conducted by observing the behavior of a single female in a petri dish arena (9 cm diameter, 2 cm height). The top of the petri dish was covered with an acrylic plate. A piece of new Sealon film (1 × 1 cm) or contaminated Sealon film used as a substratum for laying DBM eggs which contained moth scales and secretions was centered on the petri dish. A DBM egg was also placed in a part of the Sealon film described above. A mated *T. chilonis* female within 24 h after emergence which had never been exposed to host eggs was introduced into the petri dish. Then the searching path of the female was traced upon the acrylic plates with a felt pen for 15 min and the oviposition behavior was

observed. At every 15 s the tracks were marked, so that the average walking length could be computed. After 10 min of tracing, 10–33 replications were conducted for each treatment pattern. Initiation of the oviposition behavior of the female was identified by the drumming stage on a DBM egg. Experiments were carried out at $24 \pm 1^\circ\text{C}$. An opisometer was used to measure the length of the trace.

Results

The mean distance covered by walking in 15 s on the Sealon film with moth scales and secretions was significantly shorter than that on a clean Sealon film ($p < 0.01$, Kruskal-Wallis test) (Table 1). The contaminated Sealon film with or without a DBM egg was equally effective. Female wasps showed an antennal-searching behavior on the contaminated Sealon film without DBM eggs. Three typical examples of walking paths of *T. chilonis* female are shown in Fig. 1. Clearly, the behavior of the female wasp changed when she came into contact with the Sealon film contaminated with most scales and secretions.

Also *T. chilonis* females remained for a relatively longer period of time on the contaminated Sealon film than on the clean one (Table 2).

The elicitation rate of oviposition behavior after the female came into contact with a DBM egg was significantly higher on a contaminated Sealon film than on a clean one ($p < 0.01$, Fisher's exact probability test) (Table 3).

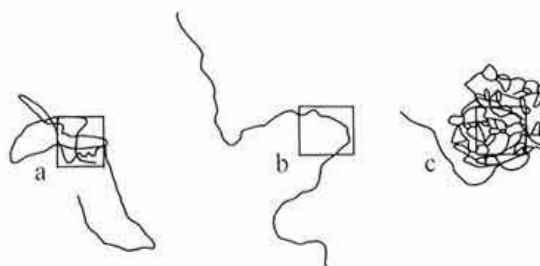


Fig. 1. Walking pattern of *T. chilonis* female in a petri dish with a central square of Sealon film
a: Film covered with moth scales and secretions without a DBM egg, b: Clean film, c: Film covered with moth scales and secretions with a DBM egg.

Table 1. Mean walking distance covered in 15 s on the Sealon film

| Presence of moth scales and secretions | Presence of DBM eggs | No. of path | Mean walking distance in 15 s Mean \pm SD (cm) |
|--|----------------------|-------------|---|
| Yes | Yes | 52 | $0.648 \pm 0.112a^*$ |
| | No | 48 | $0.638 \pm 0.116a$ |
| No | Yes | 10 | $1.631 \pm 0.369b$ |

* Values not followed by the same letter in a column are significantly different at 1% level by the Kruskal-Wallis test.

Table 2. Period of stay of *T. chilonis* female on the Sealon film

| Presence of moth scales and secretions | Presence of DBM eggs | No. of individuals | | | |
|--|----------------------|--------------------|-------|--------|------------|
| | | Period of stay | | | |
| | | <30 s | <60 s | <5 min | over 5 min |
| Yes | Yes | 3 | 1 | 1 | 9 |
| | No | 2 | 2 | 6 | 0 |
| No | Yes | 13 | 2 | 3 | 1 |

Table 3. Elicitation rate of oviposition behavior of *T. chilonis* female after coming into contact with a DBM egg

| Presence of moth scales and secretions | No. of tested females | Elicitation rate of oviposition behavior (%) |
|--|-----------------------|--|
| Yes | 17 | 90.9* |
| No | 11 | 29.4 |

* Significantly different from the elicitation rate for the absence of moth scales and secretions at 1% level (Fisher's exact probability test).

Discussion

The present results indicated that *T. chilonis* female changes its walking behavior on a Sealon film with remnants of scales and secretions of adult DBM female. It moves more slowly and changes its direction more frequently, regardless of the presence of DBM eggs (Fig. 1, Table 1), suggesting that the female wasps were searching "carefully" so that a decrease in speed may result in increased antennation per unit of space. This change is referred to as area-concentrated search or local search. Local search is characterized by a high turning rate, strong turn bias, and low locomotory activity⁵⁾. Such behavioral changes were observed in several insects including *Trichogramma* when resource or patch cues were perceived⁵⁾. *Trichogramma* has been a traditional subject in investigations on parasitoid searching behavior^{2-7,11-13,15,17,21-24,26-30)}. For example, some *Trichogramma* tended to stay in and search for other host eggs in this restricted area^{3,21,22,29)}. However, the change in the searching behavior of *T. chilonis* had not yet been studied in detail. Although it is obvious that such a behavioral switch-over is caused by the contact with moth scales and/or secretions, it remains to be determined whether contact with either moth scales or with secretions or both is actively involved.

Many studies on kairomones have been carried out in *Trichogramma*^{17,25-28)}. Although these studies showed that kairomones increased the parasitism rate of host egg, host-searching and oviposition behavior of *Trichogramma* females was not observed in detail. It remained to be determined whether the elicitation of the host-searching or oviposition behavior may account for the increased parasitism rate of the eggs. In the present study, it was shown by direct observation that moth scales and/or secretions elicited the ovipositional behavior of *T. chilonis* females on DBM eggs.

Salt²⁷⁾ considered that in *Trichogramma* a visual stimulus from the host egg was a cue to find it. However, it is assumed that the ovipositional behavior of *T. chilonis* does not occur mainly on the basis of visual stimuli, because *T. chilonis* females could not exhibit the oviposition behavior on DBM eggs present on a clean Sealon film.

Since DBM eggs are usually deposited as an egg mass, it is possible that another egg can be found near the site where the first egg was parasitized. Thus, the local search of *T. chilonis* females after

egg parasitization is considered to increase the probability for the parasitoid to encounter the next egg. Consequently switchover from faster searching to slow one induced by moth scales or secretions would be advantageous for *T. chilonis*. One problem in behavioral ecology is to determine how foraging animals exploit food distributed in patches in the environment. Iwasa et al.¹⁰⁾ showed that an optimal foraging strategy depends critically on the spatial distribution of prey. They suggested that the fixed-GUT strategy is the best strategy only when the distribution of food is highly contagious. Thus it is assumed that *T. chilonis* female shows suitable characteristics for the biological control of DBM.

References

- 1) Ankersmit, G. W. (1953): DDT-resistance in *Plutella maculipennis* Curt (Lep.) in Java. *Bull. Entomol. Res.*, **44**, 421-425.
- 2) Bai, B. B. & Smith, S. M. (1994): Patterns of host exploitation by the parasitoid wasp *Trichogramma minutum* (Hymenoptera: Trichogrammatidae) when attacking eggs of the spruce budworm (Lepidoptera: Tortricidae) in Canadian forests. *Ann. Entomol. Soc. Am.*, **87**, 546-553.
- 3) Beevers, M. et al. (1981): Kairomones and their use for management of entomophagous insects. X. Laboratory studies on manipulations of host-finding behavior of *Trichogramma pretiosum* Riley with a kairomone extracted from *Heliothis zea* (Boddie) moth scales. *J. Chem. Ecol.*, **7**, 635-648.
- 4) Bell, W. J. (1990): Searching behavior patterns in insects. *Ann. Rev. Entomol.*, **35**, 447-467.
- 5) Bell, W. J. (1991): Searching behavior: the behavioral ecology of finding resources. Chapman and Hall, London, pp. 358.
- 6) Chassain, Ch., Bouletreum, M. & Fouillet, P. (1988): Host exploitation by parasitoids: local variations in foraging behavior of females among populations of *Trichogramma* species. *Entomol. Exp. Appl.*, **48**, 195-202.
- 7) Grenier, S., Veith, V. & Renou, M. (1993): Some factors stimulating oviposition by the oophagous parasitoid *Trichogramma brassicae* Bezd. (Hym., Trichogrammatidae) in artificial host eggs. *J. Appl. Entomol.*, **115**, 66-76.
- 8) Hirashima, Y. et al. (1990): Studies on the biological control of the diamondback moth, *Plutella xylostella* (Linnaeus). 2. Effect of temperature on the development of the egg parasitoids, *Trichogramma chilonis* and *Trichogramma ostrinae*. *Sci. Bull. Fac. Agric., Kyushu Univ.*, **44**, 71-75 [In Japanese with English summary].
- 9) Iga, M. (1985): The seasonal prevalence of occurrence and the life tables of the diamondback moth, *Plutella xylostella* (L.) (Lepidoptera: Yponomeutidae).

- Jpn. J. Appl. Entomol. Zool.*, **29**, 119–125 [In Japanese with English summary].
- 10) Iwasa, Y., Higashi, M. & Yamaura, N. (1981): Prey distribution as a factor determining the choice of optimal foraging strategy. *Am. Nat.*, **117**, 710–723.
 - 11) Jennings, D. T. & Jones, R. L. (1986): Field tests of kairomones to increase parasitism of spruce budworm (Lepidoptera: Tortricidae) eggs by *Trichogramma* spp. (Hymenoptera: Trichogrammatidae). *Great Lakes Entomol.*, **19**, 185–189.
 - 12) Lewis, W. J. et al. (1975): Kairomones and their use for management of entomophagous insects. II. Mechanisms causing increase in rate of parasitization by *Trichogramma* spp. *J. Chem. Ecol.*, **1**, 349–360.
 - 13) Laing, J. (1937): Host-finding by insect parasites. I. Observations on the finding of hosts by *Alysia manducator*, *Mormoniella vitripennis* and *Trichogramma evanescens*. *J. Anim. Ecol.*, **6**, 298–317.
 - 14) Lenteren van, J. C. (1980): Evaluation of control capabilities of natural enemies; Does art have to become science? *Neth. J. Zool.*, **30**, 369–381.
 - 15) Lewis, W. J., Sparkes, A. N. & Redlinger, L. M. (1971): Moth odor: a method of host-finding by *Trichogramma evanescens*. *J. Econ. Entomol.*, **64**, 557–558.
 - 16) Liu, M. Y., Jtzeng, Y. & Sun, C. N. (1982): Insecticide resistance in the diamondback moth. *J. Econ. Entomol.*, **75**, 153–155.
 - 17) Miura, K. (1992): Biological control of the diamondback moth by the egg parasitoid. *Kagaku to Seibutsu*, **30**, 332–338 [In Japanese].
 - 18) Miura, K. & Kobayashi, M. (1993): Effect of temperature on the development of *Trichogramma chilonis* Ishii (Hymenoptera: Trichogrammatidae), an egg parasitoid of the diamondback moth. *Appl. Entomol. Zool.*, **28**, 393–396.
 - 19) Miura, K. & Kobayashi, M. (1995): Reproductive properties of *Trichogramma chilonis* females on diamondback moth eggs. *Appl. Entomol. Zool.*, **30**, 393–400.
 - 20) Miura, K., Matsuda, S. & Kobayashi, M. (1994): Discrimination between parasitized and unparasitized hosts in an egg parasitoid, *Trichogramma chilonis* Ishii (Hymenoptera: Trichogrammatidae). *Appl. Entomol. Zool.*, **29**, 317–322.
 - 21) Morrison, G. & Lewis, W. J. (1981): The allocation of searching time by *Trichogramma pretiosum* in host-containing patches. *Entomol. Exp. Appl.*, **30**, 31–39.
 - 22) Noldus, L. P. J. J. & Lenteren van, J. C. (1985): Kairomones for the egg parasite *Trichogramma evanescens* Westwood. I. Effect of volatile substances released by two of its hosts, *Pieris brassicae* L. and *Mamestra brassicae* L. *J. Chem. Ecol.*, **11**, 781–791.
 - 23) Noldus, L. P. J. J. & Lenteren van, J. C. (1985): Kairomones for the egg parasite *Trichogramma evanescens* Westwood. II. Effect of contact chemicals produced by two of its hosts, *Pieris brassicae* L. and *Mamestra brassicae* L. *J. Chem. Ecol.*, **11**, 793–800.
 - 24) Nordlund, D. A. et al. (1977): Kairomones and their use for management of entomophagous insects. VII. The involvement of various stimuli in the differential response of *Trichogramma pretiosum* Riley to two suitable hosts. *J. Chem. Ecol.*, **3**, 513–518.
 - 25) Okada, T. (1989): Parasitoids of the diamondback moth, *Plutella xylostella* (L.) (Lepidoptera: Yponomeutidae); Species and seasonal changes of parasitism in cabbage fields. *Jpn. J. Appl. Entomol. Zool.*, **33**, 17–23 [In Japanese with English summary].
 - 26) Renou, M. et al. (1992): Identification of compounds from the eggs of *Ostrinia nubilalis* and *Mamestra brassicae* having kairomone activity on *Trichogramma brassicae*. *Entomol. Exp. Appl.*, **63**, 291–303.
 - 27) Salt, G. (1935): Experiment studies in insect parasitism. III. Host selection. *Proc. Roy. Soc. ser. B. Biol. Sci.*, **117**, 413–435.
 - 28) Salt, G. (1938): Experimental studies in insect parasitism. VI. Host suitability. *Bull. Entomol. Res.*, **29**, 223–246.
 - 29) Shu, S. & Jones, R. L. (1986): Laboratory studies of the host-seeking behavior of a parasitoid, *Trichogramma nubilale* and a kairomone from its host, *Ostrinia nubilalis*. *Les Colloques de L'INRA*, **43**, 249–265.
 - 30) Weseloh, R. M. (1981): Host location by parasitoids. In *Semiochemicals—their role in pest control*, eds. Nordlund, D. A., Jones, R. L. & Lewis, W. J., Chapman & Hall, London, 79–95.

(Received for publication, January 9, 1996)