## Avian Encephalomyelitis—Review of Recent Japanese Works

## By HIKOKICHI TSUBAHARA

Chief, Viral Products Section, Biological Products Division, National Institute of Animal Health

It was thought that this disease did not exist in Japan, but since the country also saw the mass outbreak of it in 1963, the investigations and studies on this subject have been done earnestly. The outlines of these studies are mentioned here.

In the spring of 1963 Miura et al.10) discovered a case extremely similar to the disease, judging from the symptoms, and reported it to be thought as avian encephalomyelitis. However the histopathological and virological proofs could not yet be obtained. Then in the fall of 1963 there was occurrence of this disease in the imported chicks under inspection,13 and the case was histopathologically and virologically proved to be definitely avian encephalomyelitis, and received a great deal of attention as the first of the cases in Japan. After that, in 1964 and 1965 the disease occurred successively in the flocks of homeproduced chicks. 2),4),6),15),16) Since 1966 it seems that there are outbreaks of the disease everywhere, not described particularly.

The disease had its onset chiefly in chicks, less than 30 days of age. In an extreme case 100% of the chicks in a flock showed the symptoms of the disease, but the usual morbidity was less than several score %. Some of chicks, more than 50 days old, showing a symptom of weak leg, were found to be diagnosed histopathologically as avian encephalomyelitis. 15)

That the occurrence of the disease came to be seen suddenly in Japan since 1963 gives us an impression that avian encephalomyelitis first invaded this country at that time, but it

is not supported by the results of serological surveys. Miura et al.11) proved that in the blood serums collected and preserved in 1953 and 1956 there already existed those having neutralizing antibody. The survey which was conducted from 1963 to 1965 gained the results that the disease already existed in the nationwide scale. Namely Iwai5) and Miura et al.9),12) surveyed 124 breeding flocks of 32 hatcheries in 22 prefectures (Japan is divided in 46 prefectures) and reported that in each prefecture there was flocks having neutralizing antibody, and that only 11% of the tested flocks were estimated to be uninfected. Tsubahara18) also surveyed 84 breeding flocks of 39 hatcheries in 21 prefectures, obtaining much the same results. Moreover, as to flocks for egg-producing, surveyed 78 farms in 8 prefectures, 66% of them was revealed clearly to be infected. For the first invasion of encephalomyelitis in 1963 the disease is thought to spread too widely in this country.

Horiuchi<sup>2)</sup> conducted histopathological studies on the cases of both natural and experimental infections and emphasized the importance of swelling of the spinal cord nerve cells, accompanied with central tigrolysis, for differential diagnosis. Yamagiwa *et al.*<sup>20)</sup> examined changes of the nerve cells in detail, discerned that the etiologic agent of this disease chiefly attacked the large motor neurons which underwent degenerative changes and diminished reducing the number of the cells, and proposed that the disease should be called poliomyelitis. Moreover, through the

electron microscopic observations of motor neurons, they found expansion of the lumen of rough-surfaced endoplasmic reticulum of motor neurons and vacuolization of them, investigated the normal structure to diminish and change to small vacuole, and considered the relation of the phenomenon to the changes, called chromatolysis or central tigrolysis, seen by the optical microscopes.<sup>21)</sup>

For serological tests, the embryo susceptibility test and neutralization test, are being conducted using eggs, and development of a in vitro method is being tried. Sato et al.17), partially refining and concentrating the cerebral emulsion of infected chicken embryos, produced the antigen, carried out the complement fixation test using the non-heated serum of the immune bird, and obtained the result being positive. They, however, pointed out that the antigen was dubious to be applied practically because the antigen titers of cerebral emulsions varied in every lot produced. Ikeda3) conducted the agar-gel-diffusion method, using the cerebral emulsion of infected embryos as the antigen, and found a specific precipitin line to be formed between the antigen and the immune blood serum. Since if the blood serum is low in neutralization titer the reaction does not occur, there is room for further improvement in this method.

In order to demonstrate antigenic materials in the tissues of sick birds, Miyamae et al. 13),14) tried to apply the direct fluorescent antibody method; namely, they investigated the cases of both natural and experimental infections, using a fluorescent antibody preparation (dyeing titer: 320 times), which was raw gamma globulin from the immune chicken serum labelled by fluorescin isothiocyanate. In the cases showing the symptoms the brains and spinal cords were positive highly in the fluorescent antibody method, but those in the uninfected birds were negative. The birds showing no symptom among the infected flock were rarely positive in the fluorescent antibody method. Fluorescent reactions were hardly demonstrated in the viscera of infected birds. The fluorescent site in brain was the plasma

of the cells positive to Nissl's stain. It tended to difficulty to detect the specific fluorescent antigen in the sick chick that passed considerable days after onset of symptom, and so they concluded that the method was worth applying as a diagnosis in the early stage of this disease.

Miura et al.8) inoculated to normal chicks and embryos with the cerebral materials of infected chicks of which ages in day varied when the disease had its onset. In chicks, inoculated with the cerebral emulsion of a sick chick 2 days after birth, the latent period of the disease was short, the incidence high; some of inoculated embryos came to show changes after several embryo passages. On the other hand, in chicks that were inoculated with the cerebral emulsion of chick diseased 12 and 18 days after birth respectively, the latent period was long and the morbidity rate was low, and even after several passages changes in the embryos were not demonstrated. These findings proposed a problem that the invasiveness of virus would vary according to the age in day when the bird showed the symptoms.

Tsubahara et al.19) isolated the virus from a case of the disease in 1965, and investigated the pathogenicity of the virus which was from the embryos in a few passages. No abnormality was demonstrated in the infected embryos but the hatched chicks showed the symptoms several days after hatching. Baby chicks were easily infected by either inoculation intra-cerebral or oral, and the indicence was high; chicks more than 28 days old were easily infected by oral administration, but had no specific symptom such as leg weakness, tremor, and so on, and were clinicaly normal. The oral administration to the laying hens brought about temporary decrease of eggs. The virus first appeared in the liver and spleen about 5 days after oral administration, and a few days later they came to be recovered from the central nerves. In the embryos and chicks 4 days of age, the virus in the central nerve increased, but as age in day became older, the recovery of the virus came to be hard and the histopathological features of the disease were not discernible. When laying hens were administered, the virus transfered to the eggs 5~10 days after administration. These results seems to confirm that in nature this disease chiefly is seen in the chicks less than 30 days old.

For preventing of the outbreaks of this disease in younger chicks, the parental antibody plays an important part. Matsukura *et al.*<sup>7)</sup> investigated how long the antibody durated. It was cleared that, the parental antibody transferred to chicks for a long period of time after infection, and that the transferred antibody could be detected from chicks within 5 or 6 weeks old. This indicates that as long as the parent bird is immunized against the disease, the parental antibody protects the chicks during the most dangerous period when the disease has its onset.

The above-mentioned findings are the outlines of the recent studies on avian encephalomyelitis in Japan. For the present prophylaxis, live vaccine is administered the hens, 10~16 weeks of age, that are expected to become breeding birds, and the chicks are produced from the immunized breeding birds only. Thus the occurrence of the disease in the chick days is well prevented. For the virus to be used as live vaccine, utilizing the character of this virus that can hardly invade the central nerve in older chicks, the near one with those in nature is applied. However, as the propagating ability of the virus in the central nerve still remains, there is room for further improvement in this point.

## References

- Chikatsune, M., Komatsu, N., Kiuchi, A., Okamoto, T., Ebi, Y., Tsutsumi, T., Koyama, K., Kikuno, T., Ota, M., and Karasawa, S.: Infectious Disease Accompaning with Ataxia and Tremor in the Flock of Imported Chick. Jap. J. Vet. Sci., 26, 395, 1964. (Japanese Abstract)
- Horiuchi, T.: Pathological Changes of Avian Encephalomyelitis-like-disease. Jap. J. Vet. Sci., 26, 460, 1964. (Japanese Abstract)
- Ikeda, S.: Niwatori-Nosekizuien-birusu no Kanten-geru-nai-Chinköhannö (Agar gel precipitation test of avian escephalmyelitis virus). 16th Meeting of Japanese Soc. Viro-

- logy, 1968.
- Ikeda, S., Asahi, O., and Oka, M.: Outbreaks of Avian Encephalomyelitis in Chicks raised in Japan: Nat. Inst. Animal Hlth, Quart., Tokyo, 6, 127, 1966. (In English)
- Iwai, H.: Studies on a Seroepidemiological Survey of Avian Encephalomyelitis in Japan. Japan. J. Vet. Res., 15, 108, 1967. (English Summary)
- 6) Kurogi, H., Moriwaki, T., Hashiguchi, Y., Matsuda, K., and Iwashina, K.: Avian Encephalomyelitis: Outbreaks in Kyushu District and Isolation of its Virus from Affected Young Chickens: Jap. J. Vet. Sci., 28, 411, 1966 (Japanese Abstract).
- 7) Matsukura, T., Miura, S., and Kotani, T.: Niwatori-Nōsekizuien-Shizenkansenkei-Yurai-Hina ni okeru Ikō-Chūwa kōtai no Suii (Persistent period of passive neutralizing antibody in chicks derived from hens naturally infected with avian encephalomyelitis). 67th Meeting of Jap. Soc. Vet. Sci., 1969.
- Miura, S., and Miyamae, T.: Nichirei o Kotonisuru Niwatori-Nōsekizuien-Yagai-Zairo no Hina, Niwatori-Taizi ni taisuru Kibyōsei ni tsuite (Virulence of infected brains collected from diseased chickens of different ages). Virus (Japanese Jurnal of Virology), 18, 185, 1968. (Japanese Abstract)
- Miura, S., Miyamae, T., Iwai, S., and Asakura, S.: Seroepidemiological Observations on Avian Encephalomyelitis in Japan. Jap. J. Vet. Sic., 28, 440, 1966. (Japanese Abstract)
- 10) Miura, S., Miyamae, T., and Sato, G.: Virological and Immunological Studies on an Avian Encephalomyelitis-like-disease Observed in Sapporo, 1963. Jap. J. Vet. Sci., 26, 394, 1964. (Japanese Abstract)
- Miura, S., Miyamae, T., and Sato, G.: Present Status of Avian Encephalomyelitis in Japan. Jap. J. Vet. Sci., 26, 460, 1964. (Japanese Abstract)
- 12) Miura, S., Miyamae, T., Sato, G., and Ebina, T.: Hokkaido ni okeru Niwatori-Nösekizuien no Kessei-Ekigakuteki-Chōsa-Seiseki ni tsuite (Sero-epidemiological survey of avian encephalomyelitis in Hokkaido). J. Hokkaido Vet. Soc., 8, 1, 1964. (In Japanese)
- 13) Miyamae, T., Kotani, T., Matsukura, T., and Miura, S.: Niwatori-Nōsekizuien no Yagairei ni okeru Keikō-Kōtai-Chokusetsuhō no Ōyō (Diagnostic value of direct fluorescent antibody method in natural cases of avian encephalomyelitis). 67th Meeting of Jap. Soc. Vet. Sci., 1969.
- 14) Miyamae, T., and Miura, S.: Immunological Diagnosis of Avian Encephalomyelitis in

vitro II. Direct Fluorescent antibody Method. Jap. J. Vet. Sci., 30 (Supple.), 135, 1968. (Japanese Abstract)

15) Odagiri, Y., Yoshimura, S., Tomo, Y., and Inoue, T.: Avian Encephalomyelitis-likedisease of Chicken over 50 Days of Age. Jap. J. Vet. Sci., 28, 442, 1966 (Japanese Abstract)

16) Odagiri, Y., Yoshimura, S., Tomo, Y., Mochizuki, H., and Hunabashi, N.: Observations on an Avian Encephalomyelitis-like-disease. J. Japan Vet. Med. Ass., 18, 589, 1965. (in Japanese)

17) Sato, G., Watanabe, H., and Miura, S.: Immunological Diagnosis of Avian Encephalomyelitis in vitro I. Direct Complement Fixation Test. Jap. J. Vet. Sci., 30 (Supple), 134, 1968. (Japanese Abstract)

18) Tsubahara, H.: Unpublished data.

19) Tsubahara, H., Shōya, S., Harada, Y., and Sasaki, E.: Pathogenicity of a Strain of Avian Encephalomyelitis Virus Isolated in Japan. Jap. J. Vet. Sci., 30 (Supple), 133, 1968 (Japanese Abstract)

20) Yamagiwa, S., Itakura, T., and Shimizu, Y.: Shinsei-Bina no Sekizui-Kaihakushitsuen ni tsuite I. Shin-Byōmei-Teishō (Poliomyelitis in baby chicks I. Advocation of a new name). 67th Meeting of Jap. Soc. Vet. Sci., 1969.

21) Yamagiwa, S., Yamashita, T., and Itakura, T.: Shinsei-Bina no Sekizui-Kaihakushitsuen ni tsuite II. Hensei-Shinkei-Saibō no Denshi-Kenbikyō-Zō (Poliomyelitis in baby chicks II. Electron microscopic observations of degenerated nervous cells) 67th Meeting of Jap. Soc. Vet. Sci., 1969.

## Present Address of Authors

Chikatsune, Masateru: Branch Office of National Animal Quarantine, Kobe, Hyogo

Horiuchi, Teiji: National Institute of Animal Health, Kodaira, Tokyo

Ikeda, Sumio: Tohoku Branch, National Institute of Animal Health, Hichinohe, Aomori Iwai, H.: Faculty of Veterinary Medicine, Hok-

kaido University, Sapporo, Hokkaido

Kurogi, Hiroshi: Kyushyu Branch, National Institute of Animal Health, Kagoshima, Kagoshima

Matsukura, Toshihiko: Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Hokkaido

Miura, Shiro: ditto

Miyahae, Takeo: ditto

Odagiri, Yoshiharu: Faculty of Agriculture, University of Osaka Prefecture, Sakai, Osaka Sato, Gihei: Faculty of Veterinary Medicine,

Hokkaido University, Sapporo, Hokkaido

Tsubahara, Hikokichi: National Institute of Animal Health, Kodaira, Tokyo

Yamaigwa, Saburo: c/o Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Hokkaido