Mating Sequence and Evidence for the Existence of a Female Contact Sex Pheromone in *Brontispa longissima* (Coleoptera: Chrysomelidae)

Kei KAWAZU1,3, Ryoko T. ICHIKI1,2, Dung T. DANG1, and Satoshi NAKAMURA1*

1 Japan International Research Center for Agricultural Sciences (JIRCAS) (Tsukuba, Ibaraki 305-8686, Japan)
2 Biosystematics Laboratory, Graduate School of Social and Cultural Studies, Kyushu University (Nishi, Fukuoka 812-8581, Japan)

Abstract
To clarify the presence of a pheromone in *Brontispa longissima* (Coleoptera: Chrysomelidae), we observed its mating behavior and conducted a series of bioassays. When male and female *B. longissima* came in contact, the male was observed to hold and mount the female, extend its penis toward her abdominal tip, and copulate. The male was observed to hold the female only after he had touched her with his antennae and/or forelegs. Males showed similar mating attempts toward females killed by freezing. On the other hand, no males showed mating attempts toward females washed with hexane, but they did toward females washed with hexane and re-treated with hexane extract of the female body or female elytra. Furthermore, males showed similar mating attempts toward a glass dummy treated with a solvent extract of female elytra. These results indicate the presence of a female sex pheromone that is perceptible by direct contact and plays an important role in mating in *B. longissima*. When the crude extract of female elytra was subjected to silica gel chromatography, only the fraction eluted with hexane elicited mating behavior in males. Furthermore, when the active fraction was chromatographed on AgNO3-impregnated silica gel, only the fraction eluted with hexane elicited mating behavior in males. The contact sex pheromone of *B. longissima* is therefore believed to consist of one or more less-polar compounds, probably saturated hydrocarbons.

Discipline: Insect pest
Additional key words: coconut, hydrocarbon, mating behavior

Introduction

The coconut hispine beetle, *Brontispa longissima* (Gestro) (Coleoptera: Chrysomelidae), is one of the most serious pests of the coconut palm, *Cocos nucifera* L. (Arecaceae), and of several ornamental palms11,15. This beetle is thought to be native to Indonesia and Papua New Guinea, but it was accidentally introduced into Vietnam in 1999, and has been spreading rapidly to other countries, such as Thailand, the Philippines, and Hainan Province in China11,12,15. *B. longissima* inhabits the young and unopened fronds of the crown of coconut palms in all its developmental stages1. Both larvae and adults damage the leaflets of the fronds by eating the surface tissues, resulting in the russet browning of the leaves, a decrease in fruit production, and eventually withering and death11,15,21. The eggs of *B. longissima* hatch after an incubation period of about five days. The larval period is 30 to 40 days, followed by a prepupal period of three days and a pupal period of six days. The adult lives up to 220 days. The preoviposition period is one to two months20. In spite of being one of the most serious pests of the coconut palm, the ecological and physiological aspects of this insect have been little studied, and their mating behavior and how it is regulated have hitherto not been investigated.

In Southeast Asia, the coconut palm has many uses and produces economically and industrially important crops such as copra, coconut oil, and coconut shell charcoal11. In addition, coconut trees are commonly planted in coastal...
areas, nurseries, parks, and gardens to act as land cover, par-terres, and as topiary. Severe attacks by *B. longissima* lead to russet brown leaves and therefore completely disfigure palms that are grown for ornamental purposes. To protect coconut palms from infestation damage by *B. longissima*, there is an urgent need to investigate methods of controlling the populations of this beetle in Southeast Asia. However, chemical control using insecticides cannot be applied due to the beetle inhabiting the crowns of tall trees, and insecti-cides also pose risks to the environment. We have therefore been aiming to control the beetle using its pheromones. It is thought that applications of contact pheromone can raise the efficiency of the infection of filamentous fungus.

In Coleoptera, the function of the cuticular hydrocar-bons as contact sex pheromones have been extensively investigated in several taxa, such as Cerambycidae, Coccinellidae, Curculionidae, and Staphylinidae. However, among the Chrysomelidae, the presence of contact sex pheromones has been reported in only a few species: the Colorado potato beetle, *Leptinotarsa decemlineata*, the Japanese green duck leaf beetle, *Gastrophysa atrocyanea*, the blue milkweed beetle, *Chrysochus cobaltinus*, the dog-bane leaf beetle, *Chrysochus auratus*, and the mustard leaf beetle, *Phaedon cochleariae*. Although contact sex pheromones play an important role in mating behavior, they have been chemically identified in only one species, *G. atrocyanea*. Fundamental research is necessary to develop applications using synthetic pheromones; however, biological research concerning mating behavior and sex pheromones is a relatively unexplored area.

Analysis of the mating behavior of *B. longissima* will provide valuable insight into the diversity of mating systems of this beetle and assist in the development of techniques for pest control. We therefore conducted observation of the mating sequence to clarify the characteristics of the female sex pheromone in *B. longissima*.

### Materials and methods

#### 1. Insects

The *B. longissima* used in this research were obtained from colonies maintained by the National Biological Control Research Center, Kasetsart University, and the Department of Agriculture (DOA), Thailand. Since it is difficult to obtain fresh leaves of coconut plants in Japan, we instead reared the beetle colony on fresh leaves of the narrow leaf cattail, *Typha angustifolia* L. (Typhaceae), an alternative host plant of this beetle. Larvae and adults were maintained separately on fresh leaves in plastic containers (15.5 cm long x 11.5 cm side x 5.0 cm high) with a mesh window in the lid. All procedures were conducted at 25°C under a 12L:12D photoperiod and 65% relative humidity.

#### 2. Observation of mating behavior

All observations of mating behavior were carried out using an experimental arena (Fig. 1). A male and female were placed on a filter paper sheet (76 x 26 mm) surrounded by glass (thickness 2 mm, placed on the top). The mating behavior of 35 pairs of beetles was continuously observed for 30 min. The mated females and males were used. Although *B. longissima* showed multiple mating, the effect of multiple mating on the mating behavior of adults was not found in our preliminary experiments. Observation of the mating behavior of adults, 3–8 weeks after emergence, was conducted 7–11 h after lights on under 12L:12D, 25°C. In preliminary experiments, the same level of frequency in mating behavior was observed between 3-8 weeks.

#### 3. Extraction

The females were killed by freezing for 1 hr at -20°C. The elytra of the killed females were dissected and extracted...
with hexane (ca. 1 mL/female) for 1 h. After the elytra had been removed and filtrated, the crude extracts were stored at -20°C until use.

4. Chromatography

The crude extract of female elytra (50 FE: female equivalent) was poured onto a silica gel column (30mm x 5mm ID, 200 mg, Wakogel C-200, 200 mesh, Wako Pure Chemical Industries Ltd.), and successively eluted with 2 mL each of hexane, then 5%, 15%, and 50% ether in hexane and ether. The active fraction was further fractionated by column chromatography on 200 mg of silica gel impregnated with 10% AgNO₃ (30mm x 5mm ID, 200 mesh, Aldrich Chemical Company, Inc.). The column was then eluted successively with 2 mL each of hexane and 0.5%, 1%, 2%, and 5% ether in hexane. All the solvent was distilled before use.

5. Bioassay for sex pheromone

Laboratory bioassays were conducted with the experimental arena (Fig. 1). Males 3–8 weeks after emergence were bioassayed at 7–11 h after lights on under 12L:12D, 25°C. Pheromonal activity was evaluated using a black glass dummy (9.0 x 1.6 mm; 1.0 mm thickness). Color effect on male mate orientation was demonstrated in Xylotrechus pyrrhoderus⁴, Anoplophora malasiaca⁴, and holotrichia loochooana loochooana³. Since mating behavior in B. longissima was also thought to be elicited by a visual stimulus, a black dummy that resembles natural female colors was used. However, in our present experiments, the effect of the black dummy on the mating behavior in B. longissima could not be confirmed. The dummy was coated with a test solution using a glass syringe and placed at the center of the experimental arena. A male was placed nearby and allowed to make contact with it. Behavior was continuously observed for 30 min. The sexual maturity of the 3–8 weeks-old males used in the bioassay was confirmed through observation of abdominal bending behavior toward intact females. Thirty replications were performed for each bioassay.

6. Statistical analysis

Each statistical analysis was performed using JMP (version 5.0.1 J for windows, SAS Institute Inc., Cary, NC, USA 2004). Male response was analyzed using a nominal logistic regression analysis with William’s adjustment. Pairwise comparisons of male response were determined by a likelihood ratio chi-square test using contingency table analysis with the Holm adjustment.

Results and discussion

1. Mating sequence in Brontispa longissima

To clarify the existence of a pheromone in this species, the mating behavior of 35 pairs of B. longissima was observed (Fig. 2). Both females and males showed wandering behavior before the encounter, waving their antennae. Of the 35 pairs examined, 91% encountered each other within 30 minutes and showed contact behavior, in which males approached females and touched them with their antennae and/or forelegs. We present two possible explanation for the male’s approach to female within short range: 1) stimulatory volatiles and/or 2) visual stimulus. This remains to be proved in the future. After the male touched the female with his antennae and/or forelegs, he grabbed her with his forelegs (holding, 91%). The male then licked her back with his palpi, aligned his body axis with that of the female, and mounted her (mounting, 86%). Most males (63%) licked the female’s backs again during mounting behavior. The male then bent his abdomen to contact the tip of the

Observed pairs: N=35

<table>
<thead>
<tr>
<th>Contact</th>
<th>91 %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Holding</td>
<td>91 %</td>
</tr>
<tr>
<td>Mounting</td>
<td>86 %</td>
</tr>
<tr>
<td>Penis extension</td>
<td>83 %</td>
</tr>
<tr>
<td>Copulation</td>
<td>69 %</td>
</tr>
</tbody>
</table>

Fig. 2. Mating sequence in B. longissima (N = 35)

Each value indicates the percent of males exhibiting the behavior in 35 observations.
female abdomen (penis extension, 83%), and copulated successfully (copulation, 69%). Thus, we classified male behavior into five phases: contact, holding, mounting, penis extension, and copulation. Since the five phases of male behavior are conducted step by step, this mating sequence is considered to be behavior formulated with extremely high probability. Similar mating sequences were observed after encounters between males and females in several other Chrysomelidae: *L. decemlineata*<sup>9,13,16</sup>, *G. atrocyanea*<sup>19</sup>, and *P. cochleariae*<sup>5</sup>. When the male *B. longissima* attempted to mate, some females walked away. This behavior was observed when the male attempted holding and mounting behavior. This was concluded to be mate refusal behavior by female *B. longissima*.

The mating sequence of *B. longissima* appeared to start from direct contact by a male with his antenna or tarsus of the foreleg to the female antenna, pronotum, or elytra. Only when a male contacted a female with his antennae and/or tarsi were further male responses, such as holding, mounting, penis extension, and copulation, observed. Male contact with the antennae, tarsi, or palpi was considered to be involved in perception of female-specific cues. The importance of direct contact in mating behavior has been demonstrated in other Chrysomelidae: *L. decemlineata*<sup>9,13,16</sup>, *G. atrocyanea*<sup>19</sup>, *C. cobaltinus*<sup>18</sup>, and *C. auratus*<sup>18</sup>. However, more detailed examinations will be needed to determine the presence of attractant cues in the mating behavior of *B. longissima*. There still remains a possibility that other cues work in the near vicinity of individuals or for long-distance attraction in association with host plant odor.

2. Behavioral response of male *B. longissima* toward live or dead females

In the bioassays, males showed mating attempts comprising penis extension toward females that had been killed by freezing at -20°C (Fig. 3). No males showed penis extension behavior toward females that had been washed with hexane. When the washed females were re-treated with the hexane extract from a female body or female elytra, males again showed penis extension toward those females. These results indicate the presence of a female sex pheromone that is perceptible by direct contact and plays an important role.
in the mating of *B. longissima*.

3. Behavioral responses of males toward a dummy coated with female extract

When a male touched a black glass dummy coated with 1 FE of female elytra extracts, males showed holding, mounting, and penis extension behavior toward the dummy (Fig. 4). This behavior was similar to behavior toward intact females. The male response was enhanced when the dose of the extract was increased. This dose-dependent relationship indicates that mating behavior is stimulated in males by a chemical factor, i.e., a female sex pheromone. However, the percentage of male responses to the dummy with the extract tended to be smaller than to intact females (Figs. 3 and 4). There is a possibility that non-chemical, visual, acoustic, or tactile cues might play a role in sexual communication of *B. longissima* in addition to the chemical cue.

4. Behavioral responses of males toward a glass dummy coated with chromatographic fractions

Male responses toward a glass dummy coated with fractions obtained from silica gel column chromatography are shown in Figure 5. Only the non-polar hexane fraction elicited as much holding and mounting as did the crude extract. The non-polar hexane fraction and crude extract

![Graph showing behavioral responses of males toward a dummy coated with female extract](image)

**Fig. 4. Behavioral responses of males toward a dummy coated with female extract (N = 30)**

Values in the same response followed by the same letter were not significantly different according to nominal logistic regression analysis.
significantly elicited penis extension. The polar fractions, however, did not (Fig. 5), indicating that one or more non-polar compounds, probably hydrocarbons, function as contact sex pheromones in \textit{B. longissima}.

5. Behavioral responses of males toward a glass dummy coated with silica gel column chromatographic fractions

The active hexane fraction was further fractionated by column chromatography on AgNO$_3$-impregnated silica gel. The fraction eluted with hexane and crude extract significantly elicited male holding and mounting (Fig. 6). The fraction eluted with hexane elicited as much penis extension as did the crude extract, whereas ether fractions, which are considered to contain alkenes, elicited none (Fig. 6). The contact sex pheromone of \textit{B. longissima} is therefore considered to consist of one or more non-polar compounds, probably saturated hydrocarbon(s). In \textit{G. atrocyanea}, the mono-methyl alkanes 9-methyl heptacosane, 11-methyl heptacosane, 9-methyl nonacosane, 11-methyl nonacosane, 13-methyl heptacosane, 13-methyl nonacosane, 15-methyl nonacosane, and \(n\)-alkanes were identified as components of the contact sex pheromone$^{10}$. It will be necessary to elucidate the chemical structure and function of the contact pher-
omone distributed on the female body surface to understand mating behavior in *B. longissima*.

**Acknowledgments**

We would like to thank Wiwat Suasa-ard, Sopon Urachuen, Kittiya Suksen, Rut Morakote, Amporn Winotai, and Thitraporn Pundee for providing experimental devices and insects. We would also like to thank Chieko Yokokura for her technical assistance. We are grateful to Professor Sadahiro Tatsuki of the University of Tokyo and Professor Yoichi Kainoh of the University of Tsukuba for valuable suggestions on the experiment.

**References**


---

Fig. 6. Behavioral responses of males toward a glass dummy coated with AgNO₃-impregnated silica gel column chromatographic fractions (N = 30) Values in the same response followed by the same letter were not significantly different according to nominal logistic regression analysis.